REVIEW ARTICLE

Mycology

Lichens of Sri Lanka: Past discoveries, present knowledge, and future directions

B Weerakoon¹, P Wolseley², S Wijesundara³, S Nissanka⁴ and G Weerakoon^{2*}

- ¹ Postgraduate Institute of Agriculture, University of Peradeniya, Peradeniya, Sri Lanka.
- ² Algae, Fungi and Plant Division, Department of Sciences, Natural History Museum, London, UK.
- ³ National Institute of Fundamental Studies, Hantana Road, Kandy, Sri Lanka.
- ⁴ Department of Crop Science, Faculty of Agriculture, University of Peradeniya, Peradeniya, Sri Lanka.

Submitted: 03 March 2025; Revised: 21 July 2025; Accepted: 25 July 2025

Abstract: Lichens are an important component of Sri Lankan biodiversity and have contributed greatly to the tropical diversity as well as to their application as bioindicators of changing environmental conditions. This review summarises 49 selected research publications on the taxonomic and chemical diversity of lichens, their application as bioindicators, and as a source of bioactive compounds that have medicinal properties against microorganisms with the potential of curing some diseases. The chemical properties of lichens and associated fungi and mycobionts have been a major focus in recent years, and several useful compounds have been identified that have benefits for human health. Several areas of the country are well surveyed and further surveys in new areas will aim to discover new species and document lichen diversity, and responses to environmental pollution. These results highlighted the importance of ongoing research in Sri Lanka in a tropical and global lichen context.

Keywords: Bio-indicators, lichen diversity, pollution, taxonomy, tropical forests

INTRODUCTION

Different ecosystems and forest types in Sri Lanka

Sri Lanka is a continental island (7.8731° N, 80.7718° E) with an area of 65,610 km² located in the Indian Ocean, southeast of mainland India, in the tropical belt,

and has a hot and humid climate (Chandrajith, 2020; Punyawardena, 2020). The mean annual temperature is 27.5 °C in the lowlands, and 15.9 °C in the highlands. Sri Lanka has two monsoon seasons and two inter monsoon seasons receiving 900 mm - 5500 mm rain p.a. The Southwest monsoon (SWM) (May- September) brings a high rainfall to the Southwest part of the country while Northeast monsoon (NEM) (December- February) causes heavy rains throughout the country. During SWM, the highest rainfall is received by the mid-elevations of the western slopes of the highlands. During NEM period, the highest rainfall is received by north-eastern parts of the central highlands (Punyawardena, 2020). According to the topography, Sri Lanka is divided into three topographical zones; low-country (< 300 m amsl), mid-country (300-900 m amsl), and upcountry (> 900 m amsl) (Chandrajith, 2020). Following the amount of rainfall received and its distribution, Sri Lanka is divided into three climatic zones; wet zone, intermediate zone and dry zone (Punyawardena, 2020).

Within the tropical climatic belt, several forest types can be seen in Sri Lanka relating to their geographical distribution (MOE, 2012). Dry forests, lowland rainforests, sub montane forests, and montane forests are the main forest vegetation types recorded (Kathriachchi,

^{*} Corresponding author (gothamie.weerakoon2@nhm.ac.uk; phttps://orcid.org/0000-0001-8290-2910)



2012; Perera, 2012; Wijesundara, 2012). Grasslands and mangroves are the main components of the non-forest vegetation type of the country (Jayatissa, 2012; Perera, 2012).

Present understanding about lichens

A lichen is a self-sustaining ecosystem formed by the interaction of an exhabitant fungus (mycobiont) and an extracellular arrangement of one or more photosynthetic partners (photobionts) and an intermediate number of other microscopic organisms (Hawksworth & Grube, 2020). The photosynthetic partner of a lichen is an alga and/or a cyanobacterium. In addition, bacteria, secondary fungi, and endolichenic fungi can be involved in the lichen symbiosis association (Weerakoon, 2015; Lücking & Spribille, 2024). The photobionts produce carbohydrates, and the mycobiont produces the lichen structure, including protective mechanisms to maintain photobiont viability. There is a growing suggestion that the association is an obligate parasitism maintained by the mycobiont (Morillas *et al.*, 2022).

Lichens can have both sexual and asexual reproduction. Both photobiont and mycobiont may reproduce asexually within the lichen. However, sexual reproduction only occurs in the mycobiont partner. Spores and asexual thallus propagules with both symbionts can be dispersed by wind, water, or mechanical forces. After being dispersed, a new lichen thallus may establish in the new habitat, if it is environmentally suitable (Purvis, 2000; Weerakoon, 2015). According to Brunialti *et al.* (2021), species which show sexual reproduction are more widely distributed than the species with asexual vegetative reproduction. Also, they reported a nested assemblage of vegetatively reproducing lichens mainly in old growth forests, while sexually reproducing lichen species showed a high turnover.

There are about 20,000 lichen species identified throughout the world (Lücking *et al.*, 2017; Finger Lakes Land Trust, 2024), and lichenologists believe there are more species yet to be discovered (Lücking & Spribille, 2024). In Sri Lanka, the lichen flora is poorly studied. Most Sri Lankan surveys have been conducted in the central highlands and wet zone lowlands, however, the intermediate zone and the dry zone lichens are less explored (Weerakoon, 2015; Jayalal *et al.*, 2020).

Lichens are found on a wide variety of substrates including soil (terricolous), rock (saxicolous), decaying

wood (lignicolous), trees (corticolous), and leaves (foliicolous). Lichens are grouped according to the shape and morphology of the thallus (Purvis, 2000). The main types being crustose (a crust-like thallus), foliose (a leaf-like thallus), fruticose (a bush-like thallus), filamentous (a bushy thread-like thallus), and squamulose (a micro leafy thallus). Foliose, filamentous and fruticose lichens are known as macro-lichens and the remainder are referred to as micro-lichens (Purvis, 2000; Weerakoon, 2015).

In the Eastern Himalaya and Western Ghats, the majority of lichens are recorded on tree trunks and the dominant group on tree trunks is crustose (Pinokiyo et al., 2008, Vinayaka et al., 2016). Also crustose lichens are recorded as the most abundant group across all habitats in temperate and tropical regions (Nayaka & Upreti, 2005; Rout *et al.*, 2010; Dudani *et al.*, 2015). Foliose and fruticose lichens are common in well-lit canopy gaps and openings and become more common at higher elevations in tropical forests (Joshi *et al.*, 2011; Kumar *et al.*, 2011; Abas & Din, 2021; Orock & Fonge, 2022).

Research conducted in tropical regions has shown that Graphidaceae is the largest and most diverse family of crustose lichens in the tropics, and that the global hotspot for this family is South India and Sri Lanka (Lücking et al., 2014). Lichens in tropical regions become more abundant, dense, and widely scattered with increasing elevation above 1000 m (Baniya et al., 2010; Kumar et al., 2011; Costas et al., 2021; Bhagarathi et al., 2022; Orock & Fonge, 2022). Further, Pinokiyo et al. (2008) also recorded a unimodal pattern in lichen diversity in relation to altitude from a study conducted in Arunachal Pradesh in India.

Lichens produce both primary and secondary metabolites (Ren et al., 2023). Primary metabolites are used in structural growth and development of the lichen thallus by both symbionts, while secondary compounds are only produced by the fungus and have a range of protective functions in the lichen. Secondary metabolites are small, complex, water insoluble, crystalline compounds deposited on the hyphae of the fungus. There are three main pathways for synthesizing these compounds; acetyl-malonate, mevalonate, and shikimate pathways (Ren et al., 2023). There are over 1000 lichen secondary compounds identified so far (Ren et al., 2023). These compounds have been known to protect the thallus from herbivory and UV radiation (Schweiger et al., 2021), and to contribute to ecological services such as nutrient

cycling, habitat formation, and soil formation (Prokopiev et al., 2025). Lichen secondary compounds also have an active pharmaceutical role in human health as antibacterial, anti-cancer, anti-fungal, anti-inflammatory, anti-microbial, antioxidant, and wound healing agents (Moreira et al., 2015; Bhattacharyya et al., 2016; Ren et al., 2023).

Lichens are one of the commonly used bioindicators for assessing atmospheric air quality since their growth and development depend on air borne resources, thus any atmospheric changes may have an impact on them (Nimis et al., 2002 Conti, 2008; Delves et al., 2023). A significant change in lichen communities can be observed due to air pollutants such as sulphur dioxide (SO₂) from burning coal and fossil fuels, and ammonia (NH₂) from agricultural activity (Sutton et al., 2009). In temperate countries and conditions, lichen communities have been widely used to recognise the impact of atmospheric pollution. An Index of Atmospheric Purity (IAP) was developed by Leblanc and DeSloover in 1970 as a method to assess air quality using lichens as bioindicators, and has since been widely used in temperate regions (Gombert et al., 2004). Biomonitoring studies using lichens have contributed to creating regional policies such as introducing critical levels and loads of pollutants affecting the lichens, such as NH, (Sutton et al., 2009). NH, is a pollutant alkaline gas that causes an increase in the bark pH of the surrounding vegetation. Lichens that prefer a lower bark acidity (acidophytes) are eliminated from the habitat and replaced by lichens that are tolerant of the increased bark pH (Wolseley et al., 2006). Modelling studies have shown that natural habitats in South Asia, such as Himalayan forests, are threatened by increasing atmospheric nitrogen deposition (Ellis et al., 2022). Further, deposited particulate matter has been shown to affect lichen thalli close to highways (Marmor & Randlane, 2007), and lichens have been used to monitor heavy metal pollution due to Pb in urban areas (Hasairin et al., 2020).

Globally, lichens are well studied in many research fields such as ecology, environmental monitoring, air quality, and medical studies as well as having commercial and cultural values in some countries (Lücking & Spribille, 2024). However, the lichen flora in Sri Lanka is less studied compared to other countries in tropical Asia (Weerakoon *et al.*, 2019). This review summarizes the findings of selected lichenological research in Sri Lanka, the status, and the potential for lichenology in Sri Lanka.

Lichen diversity and distribution in Sri Lanka

History of lichen surveys in Sri Lanka

G.H.K. Thwaites was the director of the Botanic Gardens in Sri Lanka from 1857-1880 and made the first lichen collection in Sri Lanka, duplicates of which are deposited in the Natural History Museum in London and other herbaria in Europe (Weerakoon, 2013). Using this collection, Leighton in 1869, published 196 lichen species for Sri Lanka, including 43 species new to science. Many foreign lichenologists conducted field explorations in the country and results were published in late 19th and 20th centuries (Nylander 1866:1900; Alston, 1938; Kurokawa, 1973; Kurokawa & Mineta 1973). Under the 'Flora of Ceylon' project by the Smithsonian Institute, R. Santesson, A. Tehler and L. Wheeler (1970 -1976) studied the lichen diversity in different parts of the country. Mason Hale made a significant contribution to the lichen flora in Sri Lanka from 1976 to 1981 including 'A revision of the lichen family Thelotremataceae in Sri Lanka' in 1981 (Hale, 1981). Brunnbauer (1984 - 1987) updated the Sri Lankan lichen flora to 546 species. Lichen discoveries by Jayasuriya (1984), Moberg (1986 and 1987), Awasthi (1991), Makhija and Patwardhan (1992) and Vezda et al. (1997) brought the lichen checklist to 659 species (Weerakoon, 2015). A group of local and foreign lichenologists initiated a series of lichen workshops from 1999 onwards and continued to increase the lichen knowledge and interest of Sri Lankan researchers (Wijesundara & Karunaratne, 2015). After 2010, local experts contributed more to reporting and describing new lichen species.

Lichen species delimitation in Sri Lanka was supported by morphology and gene-based analyses conducted on specimens collected from the Central Mountain range. Studies in Knuckles showed that lichen specimens in the genus Pyrenula of family Pyrenulaceae are not derived from the same ancestor and can be divided into two groups (Weerakoon et al., 2012 a). In the Horton Plains, Jayalal et al. (2012) described two new foliose lichen species in genus Anzia. Also, morphological descriptions continued, particularly in crustose species, following Weerakoon's research in the Knuckles Mountain range (Weerakoon et al., 2012a; 2012b; 2012c; Wijeyaratne et al., 2012). Further, three new lichen species in genera Heterodermia, Malmidea, and Protoparmelia and an identification key for the genus *Heterodermia* were published for Sri Lanka (Weerakoon & Aptroot, 2013). Research on the family Graphidaceae added 13 new

species (Weerakoon *et al.*, 2014) and 6 new species from the montane forests of Horton Plains by Weerakoon *et al.* (2015). From an examination of a collection made in the early 1990's, Weerakoon & Aptroot (2014) recorded 207 lichens new to Sri Lanka including three species new to science, 91 new records for the Indian subcontinent, and four new records for Asia.

Further research by Weerakoon *et al.* (2016) and Weerakoon and Aptroot (2016) increased the number of new records for Sri Lanka to 152 species of which 17 were described as new to science, 86 records as new to the Indian subcontinent and eight as new to Asia. In the same year the first record of genus *Cora* in the eastern palaeotropics was described from the Sinharaja rain forest reserve, Sri Lanka (Lucking *et al.*, 2016). Furthermore, three new *Ocellularia* species (Family Graphidaceae) were identified from Sri Lanka (Li *et al.*, 2016). In 2018, further ten species were recorded with three species new to science by Aptroot & Weerakoon (2018). *Leightoniella zeylanensis*, an endemic species, was rediscovered and classified under family Pannariaceae using molecular analysis (Weerakoon *et al.*, 2018).

In 2019, the first record of palaeotropical *Allographa* with pigmented lirellae (Family Graphidaceae) was recorded from Sri Lanka together with a global key for all *Allographa* species with colourful lirellae. These *Allographa* species were previously recorded only from the Neotropics and from Africa (Jatnika *et al.*, 2019).

Six new species of *Allographa* and *Graphis* of Graphidaceae with 106 new records were reported with a key to genera for Sri Lanka by Weerakoon *et al.* in 2019. The total number of species of *Allographa* and *Graphis* in Sri Lanka amounted to 124 species, making Sri Lanka the hotspot for *Allographa* and *Graphis* diversity in the world. Research on genus *Phyllopsora* in Asia documented 18 species for Sri Lanka and a taxonomic key for Asian species (Kistenich *et al.*, 2019). In 2020, Kistenich *et al.* also recorded *Aciculopsora* in Sri Lanka, the first record in the paleotropics of a genus previously known only in neotropics.

With the contribution of local and foreign lichenologists, the documented lichen flora in Sri Lanka (as of 2020) stands at 876 species across 233 genera in 60 families (Jayalal *et al.*, 2020). Since then following further research on historic lichen collections from Sri Lanka, Elvebakk (2021) has described a new species of cyanolichen, *Gibbosporina cyanea*, based on the holotype collected by Thwaites in the 1860s.

Factors affecting the lichen distribution in Sri Lanka

Lichens are distributed across all climatic regions of Sri Lanka and the distribution pattern differs in each region. Environmental variables such as rainfall, humidity, and temperature affect this variation (Weerakoon, 2015). Altitude strongly affects lichen distribution and community composition (Weerakoon *et al.*, 2020) so that lichen diversity increased with altitude. As observed in mountain forests in the wet-zone (Weerakoon *et al.*, 2020), Gunawardena and Wijeyaratne, (2020) also described high lichen diversity in the Ritigala forest, an isolated mountain in the dry-zone of the country. The study also reported the influence of light exposure on lichen communities, the preference for sunny areas by lichens with a green algal photobiont and for shaded habitats by cyanobacterial lichens.

Factors that affect lichen diversity and distribution include forest disturbance (Weerakoon et al., 2010; Weerakoon et al., 2020). In the Knuckles Mountain range, Sri Lanka (in the central highlands) the highest lichen diversity was recorded in pristine forest while monoculture plantations such as tea, pine and acacia had the lowest lichen diversity (Weerakoon et al., 2010; Weerakoon et al., 2020). These studies also demonstrated that corticolous lichen diversity increased where tree diversity was high in undisturbed forests. De Silva and Senanayake (2015) recorded low lichen diversity in a monoculture pine plantation compared with adjoining secondary forest in Pussellawa in the central highlands. The lichen diversity was higher in the montane forest islands than in continuous forests in the Horton Plains National Park – the southern part of the central highlands where fire caused frequent disturbance (Jayalal et al., 2017).

Other research in Sri Lankan lichenology

Although the chemical composition of lichens has been used to define lichen species, many studies have been conducted on lichens to assess the application of lichen compounds to human health and environmental problems.

Chemical compounds of lichens and their potential applicability

The chemical properties of lichens and their uses were studied in Sri Lanka from the early 2000s (Karunaratne *et al.*, 2005) including an investigation of larvicidal assays in relation to mosquito larvae (Nanayakkara

et al., 2005). An investigation of secondary metabolites in butterfly larvae highlighted the role of secondary metabolites in protecting lichens from herbivores (Karunaratne et al., 2008). Secondary metabolites were used to define new species (Kathirgamanathar et al., 2006; Bombuwela et al., 2008) and then further developed to assess their application to human health. Several recent studies have highlighted the potential of secondary chemical compounds generated from lichen fungi in medical research. Studies have shown that lichens in the *Parmotrema* genus have antioxidant properties (Samanthi et al., 2015). Two lichen species in genus Parmotrema, P. rampoddense and P. tinctorum, showed potential importance as a source for future antimicrobial drugs (Shiromi et al., 2021). Additionally, the anti-diabetic properties of the secondary metabolites of Cladonia sp. were documented by Karunaratne et al. (2014).

Antibacterial compounds of lichen-associated fungi in mangrove habitats were identified by Happitiya et al. (2023a; 2023b). Weerasinghe et al. (2021) also tested the antioxidant, anti-inflammatory and antibacterial properties of the secondary metabolites extracted from endolichenic fungi from lichens which occurred in mangrove habitats in Negombo, Sri Lanka. The antioxidant, anti-lipase, and anti-amylase properties of the secondary metabolites in endolichenic fungi are documented together with their anti-inflammatory properties (Maduranga et al., 2018, 2021). A novel chemical compound extracted from an endolichenic fungus Amandinea medusulina found in mangrove ecosystems showed a moderate activity against human lung cancer (Santhirasegaram et al., 2020). In 2022, a chemical compound with high antibiotic properties was extracted from a mangrove associated lichen. The compound has high anti-oxidant properties, moderate anti-inflammatory properties, some activities, and moderate cytotoxicity against oral cancer. This chemical was extracted from an endolichenic fungus recorded from the host lichen Bactrospora myriadea from the Negombo lagoon mangrove community (Weerasinghe et al., 2022). Further, secondary metabolites extracted from endolichenic fungi of mangrove associated lichens Arthonia antillarum and Bactrospora myriadea have shown cytotoxicity against human breast, oral and lung cancers (Shevkar et al., 2024).

Using lichens as bio-indicators for assessing forest ecosystem health

Lichens have been used to detect changes in communities in response to changes in environmental conditions as

well as air quality, and more recently to global warming. Changes in lichen communities of forests in relation to disturbance was quantified in plots in habitats in the Knuckles Mountain range, Sri Lanka (Weerakoon, 2013). Weerakoon et al. (2020) showed that 57 out of 60 indicator lichens were serving as strong and unique indicator species for an individual vegetation type in Knuckles mountain range. Of these, the prevalence of crustose species in the genera Myriotrema, Ocellaria, and Porina was associated with the undisturbed montane and sub-montane forests in Sri Lanka (Weerakoon et al., 2010). In 2010, Jayalal also found several foliose lichen genera that were good indicators of ecological continuity in the Horton Plains National Park. These results support the use of corticolous lichens to evaluate the habitat conditions in tropical forests of Sri Lanka (Jayalal, 2010; Weerakoon, 2010; Weerakoon et al., 2010; Weerakoon et al., 2020).

Air pollution monitoring in Sri Lanka using lichens

Lichens have been used as air pollution indicators in several locations in Sri Lanka. The IAP in the Horton Plain National Park, Sri Lanka was calculated from the lichen diversity. The low concentrations of NO₂ and SO₂ facilitated a high lichen diversity in the area (Jayalal et al., 2017). Similarly, the IAP was negatively correlated with the NO, and SO, levels at different locations in Kegalle in the Sabaragamuwa province (Yatawara & Dayananda, 2019). These studies have helped to identify air pollution tolerant indicator species, such as *Pyxine* sp. A recent study was conducted as a collaborative work in Sri Lanka and India, to assess the use of lichens as air pollution indicators. A comparison of lichen diversity and air pollution status in selected locations in and around Kandy city (Sri Lanka) showed a higher lichen diversity in less air-polluted locations. The highest species richness was recorded from the Hantana forest, which was located away from the city centre, compared to the Udawatta Kele and Gannoruwa forests, which were located in and around Kandy city (Preeti et al., 2023). Further work recorded higher lichen species richness in the University of Peradeniya, a semi-disturbed area, than in the polluted area around the lake in Kandy city (Edirisinghe & Athukorala, 2024). Similarly, Gunawardena et al. (2021) recorded higher lichen species richness in the University of Peradeniya than in the Kandy city centre, where they also detected increased secondary metabolites produced as a response to pollution stress.

Atmospheric pollution from NH₃ is an emerging threat to both environmental and human health, and South Asia is considered a hotspot of NH₂ pollution. An

experimental study is ongoing in a sub-montane forest reserve as a part of the South Asian Nitrogen Hub project to assess lichen responses to the elevated NH₃ levels in order to understand real world NH₃ pollution conditions (Sutton *et al.*, 2022). The multi-layer model used in this study is applicable for identifying the fate of NH₃ in tropical forest ecosystems with special focus on lichen bio-indicators, that will provide vital evidence to inform the establishment of NH₃ critical levels and associated nitrogen policy development in Sri Lanka and the South Asian region (Deshpande *et al.*, 2024).

Lichen biomonitoring on particle deposition in Sri Lanka

Airborne particles directly impact human health. They are one of the major factors causing respiratory tract inflammations and diseases. Very few studies were conducted using lichens as bio-monitors for pollutant deposition. Assessing metal deposition on lichens is a proxy for the status of the air quality. Samples of *Heterodermia speciosa* were collected from Colombo (Western Province) and Kurunegala (North-western Province) in Sri Lanka and analysed for metal deposition. They recorded K, Ca, Ti, Fe, Mn, Zn, and Pb from analysed lichen thalli. X-ray fluorescence spectrometry shows that metal deposition is higher in Colombo than in Kurunegala, indicating the air pollution in Colombo is more than in Kurunegala (Gunathilaka *et al.*, 2011).

CONCLUSION

The rich lichen diversity in Sri Lanka has been demonstrated from previous lichenological surveys conducted in limited geographical areas in the country. Further surveys of undocumented areas of Sri Lanka will reveal new records and species to confirm Sri Lanka as a hotspot for tropical lichens, highlighting the contribution to global lichen diversity. Also, medicinal properties of lichens and their use as bioindicators have been explored in recent years. Several chemical compounds extracted from lichens have shown antibiotic, antibacterial, antimicrobial, antioxidant, anti-diabetic, anti-inflammatory, and anti-cancer properties. These chemicals compounds could be used in the pharmaceutical industry to generate income and provide benefits for human health. Further, lichens have been used as bioindicators to monitor air quality in Sri Lanka responding to atmospheric NO₂, SO₂, and metal levels in different parts of the country, especially in urban areas. Ongoing studies can be used as baseline information for proposing critical levels of air pollutants, such as NH3, for tropical regions. Also, sensitive and tolerant lichen species to changes in air quality can be proposed based on current studies. Critical levels of NH₃ are already established in countries in temperate regions, where citizen monitoring programs have been conducted to increase community engagement on monitoring air quality using bioindicators. Public awareness and engagement on lichen biomonitoring and lichen conservation should be another future direction in lichen studies in Sri Lanka.

REFERENCES

- Abas, A., & Din, L. (2021). The Diversity of Lichens along Elevational Gradients in the Tropical Montane Forest of Selangor, Malaysia. *Sains Malaysiana*, 50(4), 1199-1209. http://doi.org/10.17576/jsm-2021-5005-01
- Alston, A. H. G. (1938). Kandy Flora, Colombo, Sri Lanka.
- Aptroot, A., & Weerakoon, G. (2018). Three new species and Ten new records of Trypetheliaceae (Ascomycota) from Sri Lanka. *Cryptogramie, Mycologie*, *39*(3), 373-378. http://doi.org/10.7872/crym/v39.iss3.2018.373
- Awasthi, D. D. (1991). A key to the microlichens of India, Nepal and Sri Lanka. *Bibliotheca Lichenologica*, 40, 1-337.
- Baniya, C. B., Solhoy, T., Gauslaa, Y., & Palmer, W. (2010). The elevation gradient of lichen species richness in Nepal. *The Lichenologist*, 42(1), 83-96. http://doi.org/10.1017/ S0024282909008627
- Bhagarathi, L. K., Maharaj, G., Da Silva, P. N. B., & Subramanian, G. (2022). A review of the diversity of lichens and what factors affect their distribution in the neotropics. *GSC Biological and Pharmaceutical Sciences*, 20(3), 27-63. https://doi.org/10.30574/gscbps.2022.20.3.0348
- Bhattacharyya, S., Deep, P. R., Singh, S., & Nayak, B. (2016). Lichen secondary metabolites and its biological activity. *American journal of Pharmtech research*, 6(6), 28-44
- Bombuwela, K., Kathirgamanathar, S., Thadani, V., Jayalal, R. G. U., Adikaram, N. K. B., Wijesundara, D. S. A., Andersen, R., Wolseley, P., & Karunaratne, V. (2008). Chemistry of Heterodermia microphylla, a lichen new to Sri Lanka. Journal of the National Science Foundation Sri Lanka, 36(3), 251-252.
- Brunialti, G., Giordani, P., Ravera, S., & Frati, L. (2021). The reproductive strategy as an important trait for the distribution of lower-trunk epiphytic lichens in Old-growth vs. non-old growth forests. *Forests*, *12*, 27. https://doi.org/10.3390/f12010027
- Brunnbauer, W. (1984 1986). Die Flechten von Sri Lanka in der Literatur. *Botanische Abteilung*, Naturehistorisches Museums Wien, (in 14 teilen, als Kopien verteilt).
- Chandrajith, R. (2020). Geology and geomorphology. In: *The soils of Sri Lanka. World soils book series*. Mapa R. Eds. Springer, Cham. 13-22. https://doi.org/10.1007/978-3-030-44144-9 3
- Conti, M. E. (2008). Lichens as bioindicators of air pollution. WIT Transactions on State of The Art in Science and Engineering, 30, 111-162. http://doi.org/10.2495/978-1-84564-002-6/05

- Costas, S. M., Canton, N., & Rodriguez, J. M. (2021). The relative effect of altitude and aspect on saxicolous lichen communities at mountain summits from central-west of Argentina. *Rodriguésia*, 72: e00282020. 2021. http://dx.doi.org/10.1590/2175-7860202172064
- De Silva, C. M. S. M., & Senananyake, S. P. (2015). Assessment of epiphytic lichen diversity in pine plantations and adjacent secondary forest in Peacock Hill, Pussellawa, Sri Lanka. *International Journal of Modern Botany*, 5(2), 29-37
- Delves, J., Lewis, J. E. J., Ali, N., Asad, S. A., Chatterjee, S., Crittenden, D., Jones, M., Kiran, A., Pandey, B. P., Reay, D., Sharma, S., Tshering, D., Weerakoon, G., van Dijk, N., Sutton, M. A., Wolseley, P. A., & Ellis, C. J. (2023). Lichens as spatially transferable bioindicators for monitoring nitrogen pollution. *Environmental Pollution*, 328, 121575. https://doi.org/10.1016/j.envpol.2023.121575
- Deshpande, A. G., Jones, M. R., van Dijk, N., Mullinger, N. J., Harvey, D., Nicoll, R., Toteva, G., Weerakoon, G., Nissanka, S., Weerakoon, B., Grenier, M., Iwanica, A., Duarte, F., Stephens, A., Ellis, C. J., Vieno, M., Drewer, J., Wolseley, P. A., Nanayakkara, S., Prabashwara, T., Bealet, W. J., & Sutton, M. A. (2024). Estimation on ammonia deposition to forest ecosystems in Scotland and Sri Lanka using wind-controlled NH₃ enhancement experiments. Atmospheric Environment, 320, 120325. https://doi.org/10.1016/j.atmosenv.2023.120325
- Dudani, S. N., Nayaka, S., Mahesh, M. K., Chandran, M. D. S., & Ramachandra, T. V. (2015). Lichen diversity in the Sacred forest fragments of Central Western Ghats. *Journal of Biodiversity Management and Forestry*, 4(2). http://dx.doi.org/10.4172/2327-4417.1000139
- Edirisinghe, E. S. M., & Athukorala, A. D. S. N. P. (2024). Can lichens be indicators for air pollution monitoring in Kandy city, Sri Lanka? *Brazilian Journal of Science*, *3*(8), 117-134.
- Ellis, C. J., Steadman, C. E., Vieno, M., Chatterjee, S., Jones, M. R., Negi, S., Pandey, B. P., Rai, H., Tshering, D., Weerakoon, G., Wolseley, P., Reay, D., Sharma, S., and Sutton, M. (2022). Estimating nitrogen risk to Himalayan forests using thresholds for lichen bioindicators. *Biological Conservation*, 265, 109401. http://doi.org/10.1016/j.biocon.2021.109401
- Elvebakk, A. (2021). *Gibbosporina cyanea* (Pannariaceae), a new bipartite cyanolichen from Sri Lanka with comparisons to related paleotropical cyanogenera. *The Lichenologist*, 53, 291-298. http://doi.org/10.1017/S002428292100027X
- Finger Lakes Land Trust. https://www.fllt.org/a-closer-lookearth-clothing-discovering-the-diversity-and-life-historyof-lichens/
- Gombert, S., Asta, J., & Seaward, M. R. D. (2004). Assessment of lichen diversity by index of atmospheric purity (IAP), index of human impact (IHI) and other environmental factors in an urban area Grenoble, southeast France. *Science of the total environment*, 324, 183-199. http://doi.org/10.1016/j.scitotenv.2003.10.036
- Gunathilaka, P. A. D. H. N., Ranundeniya, R. M. N. S., Najim, M. M. M., & Seneviratne, S. (2011). A determination of air pollution in Colombo and Kurunegala, Sri Lanka, using

- energy dispersive X-ray fluorescence spectrometry on *Heterodermia speciosa. Turkish Journal of Botany*, *35*, 439-446. http://doi.org/10.3906/bot-1006-15
- Gunawardena, K. W., & Wijeyaratne, S. C. (2020). Species diversity and altitudinal preferences of lichens on selected substrata in Ritigala Strict Natural Reserve. *Journal of* the National Science Foundation Sri Lanka, 48(1), 49-56. http://dx.doi.org/10.4038/jnsfsr.v48i1.9933
- Gunawardena, W. G. D. I., Edirisinghe, E. S. M., Abayasekara, C. L., & Athukorala, A. D. S. N. P. (2021). Air pollution affects lichen species richness, species density, relative growth form abundance and their secondary metabolite production: a case study in Kandy district, Sri Lanka. *Ruhuna Journal of Science*, 12(2), 115-127. http://doi.org/10.4038/rjs.v12i2.106
- Hale, M. E. (1981). A revision of the lichen family Thelotremataceae in Sri Lanka. Bulletin of the British Museum (Natural History), 8, 227-332.
- Happitiya, H. A. D. N. N., Nanayakkara, C. M., Ariyawansa, K. G. S. U., Ediriweera, S. S., Wijayawardene, N. N., Jayasinghe, R. P. P. K., Don-Qin, D., & Karunarathna. S. C. (2023 a). Antibacterial activities of lichen-associated fungi in mangrove ecosystems in Sri Lanka as potent candidates for novel antibiotic agents. In: Proceedings of SLIIT International Conference on Advancements in Sciences and Humanities, 1-2 December, Colombo, 397-403.
- Happitiya, H. A. D. N. N., Nanayakkara, C. M., Ariyawansa, K. G. S. U., Ediriweera, S. S., Wijayawardene, N. N., Jayasinghe, R. P. P. K., Don-Qin, D., & Karunarathna. S. C. (2023 b). Lichen- associated fungi inhabiting from a mangrove ecosystem in Sri Lanka: A novel source of antibacterial agents. SLIIT Journal of Humanities and Sciences. 4(2), 40-49. http://doi.org/10.4038/ajhs.v4i2.58
- Hasairin, A., Pasaribu, N., & Siregar, R., (2020). Accumulation of Lead (Pb) in the lichen thallus of mahogany trees in Medan city road. Water Air Soil Pollut, 231, 256. https:// doi.org/10.1007/s11270-020-04625-8
- Hawksworth, D. L., & Grube, M. (2020). Lichens redefined as complex ecosystems. *New Phytologist*, 227, 1281-1283.
- Jatnika, M. F., Weerakoon, G., Arachchige, O., Noer, I. S., Voytsekhovich, A., & Lücking, R. (2019). Discoveries through social media and in your own backyard: two new species of *Allographa* (Graphidaceae) with pigmented lirellae from the paleotropics, with a world key to species of this group. *The Lichenologist*, 51(3), 227-233. http://doi. org/10.1017/S0024282919000094
- Jayalal, R. G. U. (2010). Study of Diversity and Taxonomy of lichens in the Horton Plains National Park with a view to biomonitor the ecosystem health. *Doctor of Philosophy* thesis, University of Peradeniya
- Jayalal, U., Wolseley, P., Gueidan, C., Aptroot, A., Wijesundara, S., & Karunarathne, V. (2012). Anzia mahaeliyensis and Anzia flavotenuis, two new lichen species from Sri Lanka. The Lichenologist, 44(3), 381-389. http://doi.org/10.1017/ S0024282911000946
- Jayalal, R. G. U., Ileperuma, O. A., Wolseley, P., Wijesundara, D. S. A., & Karunaratne, V. (2017). Correlation of atmospheric purity index to the diversity of lichens in the Horton plains

national park, Sri Lanka. Ceylon Journal of Science, 46(2), 13-29. http://doi.org/10.4038/cjs.v46i2.7426

- Jayalal, U., Weerakoon, G., Wolseley, P., Wijesundara, S., & Karunaratne, V. (2020). A provisional list of lichens in Sri Lanka. In: The National Red List 2020- Conservation status of the flora of Sri Lanka. Biodiversity Secretariat of the Ministry of Environment and the National Herbarium, Department of National Botanic Gardens, Peradeniya, 214-225.
- Jayasuriya, A. H. M. (1984). Flora of Ritigala Nature Reserve, Sri Lanka. Forester, XVI(3 &4).
- Jayatissa, L. P. (2012). Present Status of Mangroves in Sri Lanka In: The National Red List 2012 of Sri Lanka; Conservation Status of the Fauna and Flora. Weerakoon, D.K. & S. Wijesundara Eds., Ministry of Environment, Colombo, Sri Lanka. 197-199.
- Joshi, S., Upreti, D. K., & Das, P. (2011). Lichen diversity assessment in Pindari Glacier Valley of Uttarakhand, India. *Geophytology*, 41(1-2), 25-41
- Karunaratne, V., Bombuwela, K., Kathirgamanathar, S., & Thadhani, V. M. (2005). Lichens: a chemically important biota. *Journal of the National Science Foundation* Sri Lanka, 33(3), 169-186
- Karunaratne, V., Kathirgamanathar, S., Wijesekera, A., Wijesundara, D. S. A., & Wolseley, P. (2008). Insight into the unique butterfly - lichen association between *Talicada* nyseus nyseus and *Leproloma sipmanianum*. Journal of Plant interactions, 3(1), 25-30. http://doi.org/10.1080/174 29140701740061
- Karunaratne, V., Thadhani, V. M., Khan, S. N., & Choudhary, M. I. (2014). Potent α-glucosidase inhibitors from the lichen *Cladonia* species from Sri Lanka. *Journal of the National Science Foundation Sri Lanka*, 42(1), 95-98. http://dx.doi.org/10.4038/jnsfsr.v42i1.6684
- Kathirgamanathar, S., Wickramasinghe, A., Bombuwela, K., Wolseley, P., & Karunaratne, V. (2006). Chemistry of two new Leprarioid lichens from Sri Lanka. *Journal of the National Science Foundation Sri Lanka*, 34(2), 85-90.
- Kathriarachchi, H. S. (2012). Present status of Lowland Wet
 Zone Flora of Sri Lanka. In: The National Red List 2012
 of Sri Lanka; Conservation Status of the Fauna and Flora.
 Weerakoon, D.K. & S. Wijesundara Eds., Ministry of
 Environment, Colombo, Sri Lanka. 175-180.
- Kistenich, S., Bendiksby, M., Vairappan, C. S., Weerakoon, G., Wijesundara, S., Wolseley, P. A., & Timdal, E. (2019). A regional study of the genus Phyllopsora (Ramalinaceae) in Asia and Melanesia. *MycoKeys*, 53, 23-72. http://doi. org/10.3897/mycokeys.53.33425
- Kistenich, S., Bendiksby, M., Weerakoon, G., & Timdal, E. (2020). A revision of the genus *Aciculopsora* (Ramalinaceae), with the description of one new species and one new combination. *Plant and Fungal Systematics*, 65(1), 200-209. https://doi.org/10.35535/pfsyst-2020-0015
- Kumar, R. S., Thajuddin, N., & Upreti, D. K. (2011). Diversity of Lichens in Kollihills of Tamil Nadu, India. *International Journal of Biodiversity and Conservation*, 3(2), 36-39
- Kurokawa, S., (1973). Supplementary notes on the genus *Anaptychia. Journal of the Hattori Botanical Laboratory*.

- 37, 563-607.
- Kurokawa, S., & Mineta, M. (1973). Enumeration of Parmeliae of Ceylon. Ann. Rept. Noto Marine Lab., Univ. Kanazawa, *13*, 71-76.
- Leighton, W. A. (1869). The Lichens of Ceylon, collected by G. H. K. Thwaites. *Trans. Linn. Soc. London*, 27, 161-185.
- Li, G. J., Hyde, K. D., Zhao, R. L., Hongsanan, Abdel-Aziz, F. A., Abbel-Wahab, M. A., Alvarado, P., Alves-Silva, G., Ammirati, J. F., Ariyawansa, H. A., Baghela, A., Bahkali, A. H., Beug, M., Bhat, D. J., Bojantchev, D., Boonpratuang, T., Bulgakov, T. S., Camporesi, E., Boro, M. C., Ceska, O., Chakraborty, D., Chen, J. J., Chethana, K. I. et al., (2016). Fungal Diversity Notes 253-366: taxonomic and phylogenetic contribution to fungal taxa. Fungal diversity. http://doi.org/10.1007/s13225-016-0366-9
- Lücking, R., Forno, M. D., Moncada, B., Coca, L. F., Vargas-Mendoza, L. Y., Aptroot, A., Arias, L. J., Besal, B., Bungartz, F., Cabrera-Amaya, D. M., Cáceres, M. E. S., Chaves., J. L., Eliasaro, S., Gutiérrez, M. C., Marin, J. E. H., Herrera-Campos, M. d. A., Holgado-Rojas, M. E., Jonitz, H., Kukwa, M., Lucheta, F., Madriñán, S., Marcelli, (2016). Turbo-taxonomy to assemble a megadiverse lichen genus: seventy new species of *Cora* (Basidiomycota: Agaricales: Hygrophoraceae), honouring David Leslie Hawksworth's seventieth birthday. *Fungal Diversity*. 84, 139-207. http://doi.org/10.1007/s13225-016-0374-9
- Lücking, R., Hodkinson, B. P., & Leavitt, S. D. (2017). The 2016 classification of lichenized fungi in the Ascomycota and Basidiomycota-Approaching one thousand genera. *The Bryologist*, 119(4), 361-416. https://doi.org/10.1639/0007-2745-119.4.361
- Lücking, R., & Spribille, T. (2024). The lives of Lichens: A natural history. Princeton and Oxford: Princeton University Press.
- Lücking, R., Johnston, M. K., Aptroot, A., Kraichak, E., Lendemer, J. C., Boonpragob, K., Cáceres, M. E. S., Ertz, D., Ferraro, L. I., Jia, Z., Kalb, K., Mangold, A., Manoch, L., Mercado-Díaz, J. A., Moncada, B., Mongkolsuk, P., Papong, K. B., Parnmen, S., Peláez, R. N., Poengsungnoen, V., Plata, E. R., Saipunkaew, W., Sipman, H. J. M., Sutjaritturakan, J., Broeck, D. V., Konrat, M. V., Weerakoon, G., & Lumbsch, H. T. (2014). One hundred and seventy-five new species of Graphidaceae: closing the gap or a drop in the bucket? *Phytotaxa*, 189(1), 7-38. http://dx.doi.org/10.11646/phytotaxa.189.1.4
- Maduranga, K., Attanayake, R. N., Santhirasegaram, S., Weerakoon, G., & Paranagama P. A. (2018). Molecular phylogeny and bioprospecting of Endolichenic Fungi (ELF) inhabiting in the lichens collected from a mangrove ecosystem in Sri Lanka. *PLoS ONE*, *13*(8): e0200711. https://doi.org/10.1371/journal.pone.0200711
- Maduranga, H. A. K., Weerasinghe, W. R. H., Attanayake,
 R. N., Santhirasegaram, S., Shevkar, C. D., Kate, A.
 S., Weerakoon, G., Samanthi, K. A. U., Kalia, K.,
 & Paranagama, P. A. (2021). Identification of novel bioactive compounds, Neurosporalol 1 and 2 from an endolichenic fungus, Neurospora ugadawe inhibited in the lichen host, Graphis tsunodae Zahlbr, from mangrove

- ecosystem in Puttalam lagoon, Sri Lanka. *Asian Journal of Chemistry*, *33*(6), 1425-1432. https://doi.org/10.14233/ajchem.2021.23229
- Makhija, U., & Patwardhan, P. G. (1992). Nomenclatural notes on some species of Trypethelium. *International Journal of Mycology and Lichenology*, 5(3), 237-251.
- Marmor, L., & Randlane, T. (2007). Effects of road traffic on bark pH and epiphytic lichens in Tallinn. Folia Cryptog. Estonica, Fasc. 43, 23-37
- Moberg, R. (1986). Rolfidium, a new lichen genus from Sri Lanka. *Lichenologist*, 18(4), 305-307
- Moberg, R. (1987). Lichens selecti exsiccate Upsalensis- Fasc. 2(Nos 26-50), *Thunbergia*, 5. 1-9.
- MOE (2012). The National Red List 2012 of Sri Lanka; Conservation Status of the Fauna and Flora. Ministry of Environment, Colombo, Sri Lanka.
- Moreira, A. S. N., Braz-Filho, R., Mussi-Dias, V., & Vieira, I. J. C. (2015). Chemistry and biological activity of *Ramalina* lichenized fungi. *Molecules*, 20, 8952-8987. http://doi.org/10.3390/molecules20058952
- Morillas, L., Roales, J., Cruz, C., & Munzi, S. (2022). Lichen as multi partner symbiotic relationships. *Encyclopedia*, 2, 1421-1431. https://doi.org/10.3390/encyclopedia2030096
- Nanayakkara, C., Bombuwela, K., Kathirgamanathar, S., Adikaram, N. K. B., Wijesundara, D. S. A., Hariharan, G. N., Wolseley, P., & Karunaratne, V. (2005). Effect of some lichen extracts from Sri Lanka on larvae of Aedes aegypti and the fungus Cladosporium cladosporioides. Journal of the National Science Foundation Sri Lanka, 33(2), 147-149
- Nayaka, S., & Upreti, D. K. (2005). Status of Lichen diversity in Western Ghats, India. Sahyadri E-News, Western Ghats Biodiversity Information System - Issue XVI
- Nimis, P. L., & Purvis, O. W. (2002). Monitoring lichens as indicators of pollution. P. L. Nimis, C. Scheidegger and P. A. Wolseley (eds.), *Monitoring With Lichens*, Kluwer Academic Publishers, Netherlands. 7-10.
- Nylander, W. (1866). Prodromi lichenographiae Scandinaviae Sulementia. Lichens Laoniae Orientalis. *Not. Sallsk. F. FI. Fenn. Forhadl.* (nova ser) *5*, 99-152.
- Nylander, W. (1900). Lichens Ceylonenses et Additamentum ad Lichens Japoniae. *Acta Soc. Sci. Fennicae*, 26, 1-33.
- Orock, A. E., & Fonge, B. A. (2022). Diversity of lichens at Mount Cameroon, south west region, Cameroon. *International Journal of Biodiversity and Conservation*, 14(2), 72-93. http://doi.org/10.5897/LIBC2021.1517
- Perera, A. (2012). Present Status of Dry-zone Flora in Sri Lanka. In: The National Red List 2012 of Sri Lanka; Conservation Status of the Fauna and Flora. Weerakoon, D.K. & S. Wijesundara Eds., Ministry of Environment, Colombo, Sri Lanka. 165-174.
- Pinokiyo, A., Singh, K. P., & Singh, J. S. (2008). Diversity and distribution of lichens in relation to altitude within a protected biodiversity hot spot, north-east India. *The Lichenologist*, 40(1), 47-62. http://doi.org/10.1017/S0024282908007214
- Preeti, K., Tejhani, M., Pandey, V., Dutta, V, Das, P., Weerakoon, B., Chatterjee, S., Ranasinghe, H., & Nissanka, S. (2023).

- Impacts of Pollution on Tropical Montane and Temperate Forests of South Asia: Preliminary Studies by Postgraduate Students in India and Sri Lanka. In: *Ecosystem and Species Habitat Modeling for Conservation and Restoration* (eds. Dhyani, S., Adhikari, D., Dasgupta, R., Kadaverugu, R), 355-372, Springer, Singapore. https://doi.org/10.1007/978-981-99-0131-9 19
- Prokopiev, I. A., Sazanova, K. V., Sleptsov, I. V., Filippova, G. V., Kuzmina, N. P., Frolova, D. A., & Zholobova, Z. O. (2025). Effect of Secondary metabolites of lichens on microbial communities in permafrost forest soils. *Contemporary Problems of Ecology*, 18(1), 82-100.
- Punyawardena. B. V. R. (2020). Climate. In: The soils of Sri Lanka. World soils book series. Mapa R. Eds. Springer, Cham. 13-22. https://doi.org/10.1007/978-3-030-44144-9 2
- Purvis, W. (2000). *Lichens*. Natural History Museum, London Ren, M., Jiang, S., Wang, Y., Pan, X., Pan, F., & Wei, X. (2023). Discovery and excavation of lichen bio active natural products. *Frontiers in Microbiology*, 14, 1177123. http://doi.org/10.3389/fmicb.2023.1177123
- Rout, J., Das, P., & Upreti, D. K. (2010). Epiphytic lichen diversity in a reserve forest in southern Assam, northern India. *Tropical Ecology*, 51(2), 281-288
- Samanthi, K. A. U., Wickramaarachchi, S., Wijerathne, E. M. K., & Paranagama, P. A. (2015). Two new antioxidant active polyketides from *Penicillium citrinum*, an endolichenic fungus isolated from Parmotrema species in Sri Lanka. *Journal of the National Science Foundation Sri Lanka*, 43(2), 119-126
- Santhirasegaram, S., Wickramarachchi, S. R., Attanayake, R. N., Weerakoon, G., Samarakoon, S., Wijeratne, K., & Paranagama, P. A. (2020). A novel cytotoxic compound from the endolichenic fungus, *Xylaria psidii* inhibiting the lichen, *Amandinea medusulina*. *Natural Product Communications*, 15(7), 1-8. http://doi.org/10. 1177/ 1934 578X 20933017
- Schweiger, A. H., Ullmann, G. M., Nurk, N. M., Triebel., D., Schobert, R., & Rambold, G. (2021). Chemical properties of key metabolites determine the global distribution of lichens. *Ecological Letters*, 25, 416- 426. http://doi. org/10.1111/ele.13930
- Shevkar, C., Weerasinghe, R., Dubey, G., Attanayake, R. N., Weerakoon, G., Kalia, K., Paranagama, P., & Kate, A. S. (2024). Bioprospecting of endolichenic fungus *Phanerochaeta chrysosporium* from mangrove associated lichen *Bactrospora myriadea* for anticancer leads. *Indian Journal of Microbiology*. http://doi.org/10.1007/s12088-024-02424-1.
- Shiromi, P. S. A. I., Hewawasam, R. P., Jayalal, R. G. U., Rathnayake, H., Wijayaratne, W. M. D. G. B., & Wanniarachchi, D. (2021). Chemical composition and antimicrobial activity of two Sri Lankan lichens, Parmotrema rampoddense, and Parmotrema tinctorum against methicillin-sensitive and methicillin-resistant Staphylococcus aureus. Evidence- Based Complementary and Alternative Medicine, 9985325. https://doi.org/10.1155/2021/9985325

- Sutton, M. A., Wolseley, P. A., Leith, I. D., van Dijk, N., Tang, Y. S., James, P. W., Theobald, M. R., & Whitfleld, C. (2009). Estimation of ammonia critical level for epiphytic lichens based on observations at farm, landscape and national scales, *Atmospheric Ammonia* (Sutton, M.A., Reis, S and Baker, S. M. H. eds), Springer, 71-86.
- Sutton, M. A., Wolseley, P. A., van Dijk, N., Deshpande, A. G., Dragosits, U., Twigg, M., Tang, Y. S., Braban, C., Levy, P., Bealey, W. J., Weerakoon, G., Jiang, J., Stevenson, D. S., Weerakoon, B., Nissanka, S. P., & Jones, M. R. (2022). 15 years on: Rationale and reflection on the 2006 Edinburgh Ammonia workshop in the light of emerging evidence. In: TEXTE Review of internationally proposed critical levels for ammonia- conference proceedings. German Environment Agency, 50-60.
- Vezda, A., Brunnbauer, W., & Breuss, O. (1997). Foliicole Flechten aus Sri Lanka. Annalen des Naturhisrorischen Museums in Wien, 99B, 737-742.
- Vinayaka, K. S., Chetan, H. C., & Mesta, A. R. (2016). Diversity and distribution patterns of Lichens in the midelevation wet evergreen forest, South Western Ghats, India. *International Journal of Research Studies in Biosciences*, 4(1), 15-20
- Weerakoon, G. S. K., Somaratne, S., Wolseley, P. A., & Wijeyaratne, S. C. (2010). Corticolous lichens as indicators of different forest management practices in the Dotalugala-Knuckles mountain range, In: Proceedings of the 15th International Forestry and Environment Symposium, 26-27 November 2010, University of Sri Jayawardenapura, Sri Lanka. 182-189
- Weerakoon, G., Aptroot, A., Lumbsch, H. T., Wolseley, P. A., Wijeyaratne, C., & Gueidan, C. (2012 a). New molecular data on Pyrenulaceae from Sri Lanka reveal two well-supported groups within this family. *The Lichenologist*, 44(5), 639-647. http://doi.org/10.1017/S0024282912000333
- Weerakoon, G., Wijeyaratne, S. C., Wolseley, P. A., Plata, E. R., Lücking, R., & Lumbsch, H. T. (2012 b). Six new species of Graphidaceae from Sri Lanka, *The Bryologist*, 115(1), 74-83. http://doi.org/10.1639/0007-2745-115.1.74
- Weerakoon, G., Rivasplata, E., Lumbsch, T., & Lücking, R. (2012 c). Three new species of *Chapsa* (lichenized Ascomycota: Ostropales: Graphidaceae) from tropical Asia. *The Lichenologist*, 44(3): 373-379. http://doi.org/10.1017/ S0024282911000892
- Weerakoon, W. M. G. S. K. (2013). Some environmental factors influencing diversity of corticolous lichens in selected disturbed and undisturbed vegetation types in Knuckles Mountain range in Sri Lanka. *Doctor of Philosophy thesis*, University of Sri Jayawardenapura
- Weerakoon, G., & Aptroot, A. (2013). Some new lichen species from Sri Lanka, with a key to the genus *Heterodermia* in Sri Lanka. *Cryptogamie, Mycologie*, *34*(4). 321-328. http://doi.org/10.7872/crym.v34.iss4.2013.321
- Weerakoon, G., & Aptroot, A. (2014). Over 200 new lichen records from Sri Lanka, with three new species to the science, *Cryptogamie, Mycologie*, 35(1): 51-62
- Weerakoon, G., Lücking, R., & Lumbsch, H. T. (2014). Thirteen new species of Graphidaceae (lichenized Ascomata:

- Ostropales) from Sri Lanka, *Phytotaxa 189*(1): 331-347. http://dx.doi.org/10.11646/phytotaxa.189.1.24
- Weerakoon, G. (2015). Fascinating lichens in Sri Lanka. Dilmah tea, Colombo, Sri Lanka
- Weerakoon, G., Jayalal, U., Wijesundara, S., Karunaratne, V., & Lücking, R. (2015). Six new Graphidaceae (lichenized Ascomata: Ostropales) from Horton plains National Park, Sri Lanka. Nova Hedwigia, http://doi.org/10.1127/nova_ hedwigia/2015/0241
- Weerakoon, G., Wolseley, P. A., Arachchige, O., Caceres, M. E. S., Jayalal, U., & Aptroot, A. (2016). Eight new lichen species and 88 new records from Sri Lanka. *The Bryologist*, 119(2), 131-142. http://doi.org/10.1639/0007-2745-119.2.131
- Weerakoon, G., & Aptroot, A. (2016). Nine new lichen species and 64 new records from Sri Lanka. *Phytotaxa*, 280(2), 152-162. http://dx.doi.org/10.11646/phytotaxa.280.2.5
- Weerakoon, G., Aptroot, A., Wedin, M., & Ekman, S. (2018). Leightoniella zeylanensis belongs to the Pannariaceae. Nordic Journal of Botany. e01880. http://doi.org/10.1111/njb.01880
- Weerakoon, G., Aptroot, A., Lücking, R., Arachchige, O., & Wijesundara, S. (2019). *Graphis* and *Allographa* (lichenized Ascomata: Graphidaceae) in Sri Lanka, with six new species and a biogeographical comparison investigating a potential signature of the 'biotic ferry' species interchange. *The Lichenologist*, 51(6), 515-559. http://doi.org/10.1017/S0024282919000392
- Weerakoon, G., Wolseley, P., Will-Wolf, S., & Wijeyaratne, C. (2020). Corticolous lichen species as indicators of disturbed/undisturbed vegetation types in the central mountains of Sri Lanka. *The Lichenologist*, 52, 233-245. http://doi.org/10.1017/S0024282920000109
- Weerasinghe, R. H., Maduranga, K., Attanayake, R. N., Shevkar, C., Kate, A. S., Weerakoon, G., Kalia, K., & Paranagama, P. A. (2021). Bioactive properties and metabolite profiles of endolichenic fungi in mangrove ecosystem of Negombo lagoon, Sri Lanka. *Natural Product Communications*, 16(10), 1-14. http://doi.org/10.1177.1934578X211048652.
- Weerasinghe, R. H., Shevkar, C. D., Maduranga, K., Pandey, K. H., Attanayake, R. N., Kate, A. S., Weerakoon, G., Behera, S. K., Kalia, K. S., & Paranagama, P. A. (2022).
 Bioprospecting of an endolichenic fungus *Phanerochaete sordida* isolated from mangrove associated lichen *Bactrospora myriadea*. *Hindawi Journal of Chemistry*, 22. https://doi.org/10.1155/2022/3193689
- Wijesundara, S. (2012). Present Status of Montane Forests in Sri Lanka. In: The National Red List 2012 of Sri Lanka; Conservation Status of the Fauna and Flora. Weerakoon, D.K. & S. Wijesundara Eds., Ministry of Environment, Colombo, Sri Lanka. 181-185.
- Wijesundara, S., & Karunaratne, V. (2015). Third National Workshop on Lichens: over fifteen years of progress in lichen research in Sri Lanka. *Journal of the National Science Foundation Sri Lanka*, 43(2), 195-196.
- Wijeyaratne, S. C., Lücking, R., & Lumbsch, T. (2012). Three new crustose lichen species from Sri Lanka. *Nova Hedwigia*, 94(1-2), 367-372. http://doi.org/10.1127/0029-

5035/2012/0008

Wolseley, P. A., Theobald, M. R., & Sutton, M. A. (2006). Detecting changes in epiphytic lichen communities at sites affected by atmospheric ammonia from agricultural sources. *The Lichenologist*, 38(2), 161-176. http://doi.org/10.1017/S0024282905005487

Yatawara, M., & Dayananda, N. (2019). Use of corticolous lichens for the assessment of ambient air quality along rural-urban ecosystems of tropics: a study in Sri Lanka. *Environmental Monitoring and Assessment*, 191: 179. https://doi.org/10.1007/s10661-019-7334-2