Search for Biologically Active Compounds from Sri Lankan Plants

U.L.B. Jayasinghe^{1,*} and Y. Fujimoto²

¹Institute of Fundamental Studies, Hantana Road, Kandy, Sri Lanka, ²Department of Chemistry and Materials Science, Tokyo Institute of Technology, Meguro, Tokyo 152-8551, Japan

Abstract: In a continuation of our studies towards the discovery of biologically active compounds from Sri Lankan plants, recently we have chemically investigated the various parts of *Diploclisia glaucescens* (Menispermaceae), *Filicium decipiens*, *Pometia eximia* (Sapindaceae), *Artocarpus nobilis* (Moraceae) and *Bridelia retusa* (Euphorbiaceae). These work led to the isolation of a number of ecdysones, triterpenes, saponins, chalcones, stilbenes, flavonoids including over twenty-five new natural products. Some of these compounds showed high molluscicidal, insecticidal, antifungal activities and radical scavenging properties towards DPPH.

INTRODUCTION

The flora of Sri Lanka comprises about 3500 flowering plants of which about 850 species are endemic to the island [1]. Among these, approximately 750 species are claimed to have use in the indigenous system of medicine [2]. In a continuation of our studies towards the discovery of biologically active substances from Sri Lankan plants, recently we have chemically investigated the various parts of *Diploclisia glaucescens* (Menispermaceae), *Filicium decipiens*, *Pometia eximia* (Sapindaceae), *Artocarpus nobilis* (Moraceae) and *Bridelia retusa* (Euphorbiaceae). These works led to the isolation of a number of ecdysteroids, triterpenes, saponins, chalcones, stilbenes, flavonoids including over twenty-five new natural products. Some of these compounds showed high molluscicidal, insecticidal, antifungal and antioxidant activities.

Diploclisia glaucescens (Bl.) Diels (=Cocculus macrocarpus W. & A.) is a liana of the family Menispermaceae, growing in the mid-country regions of South India and Sri Lanka. The leaves have been used in the treatment of biliousness and venereal diseases [3]. Earlier studies on the seeds of this plant described the isolation of a principle ecdysteroid, 20-hydroxyecdysone (ecdysterone) (1) (0.46% from seeds) along with four additional ecdysteroids, which were active to larvae of the European corn borer, Ostrinia nubialis [4]. 20-Hydroxyecdysone (0.5% from roots) was isolated also from the roots of D. glaucescens [5]. Our previous studies on the stem of this plant furnished 20-hydroxyecdysone in 3% yield, which is the highest recorded for the isolation of 1 from plants [6], together with a proaporphine alkaloid stepharine [7] and triterpenoids serjanic acid (2), phytolaccagenic acid (3) and a series of their glycosides [8-11].

Recently we have carried out detailed analysis on the constituents of the fruits and leaves of *D. glaucescens*. Chromatographic separation of the methanol extract of the fruits of *D*.

^{*}Corresponding author: Fax: 0094-81-2232131; E-mail: lalith@ifs.ac.lk

glaucescens furnished a new ecdysteroid, 2-deoxy-5, 20-dihydroxyecdysone (4) together with 3-deoxy-1, 20-dihydroxyecdysone (5), 2-deoxy-20-hydroxyecdysone (6), 24-ethyl-20hydroxyecdysone (makisterone C) (7) [12], bidesmosidic triterpenoidal saponins, 3-O- -Dglucopyranosylphytolaccagenic acid 28-O- -D-glucopyranosyl ester (8), 3-O-[-Dglucopyranosyl-(1 3)- -D-glucopyranosyl]phytolaccagenic acid 28-O- -D-glucopyranosyl ester (9), $3 - O - \begin{bmatrix} -L - rhamnopyranosyl - (1 2) - -D - glucopyranosyl - (1 2) - -D - glucopy$ glucopyranosyl]phytolaccagenic acid 28-O- -D-glucopyranosyl ester (10), 3-O-[-Lrhamnopyranosyl-(1 2)- -D-glucopyranosyl-(1 2)- -D-glucopyranosyl]serjanic acid 28-O- -D-glucopyranosyl ester (11) [13], 3-O-[-D-glucopyranosyl-(1 2)- -D-glucopyranosyl]serjanic acid 28-O- -D-glucopyranosyl ester (12), 3-O-[-D-xylopyranosyl-(1 2)- -D-glucopyranosyl-(1 2)- -D-glucopyranosyl]serjanic acid 28-O- -D-glucopyranosyl ester (13), 3-O- -D-glucopyranosyl-20-hydroxyecdysone (14) [14], and phenyl glycosides 4-[-D-xylopyranosyl-(1 6)- -D-glucopyranosyloxy]benzonitrile (15) and 4-(2-nitroethyl)phenyl -D-xylopyranosyl -(1 6)- -D-glucopyranoside (16) [15]. Chromatographic separation of the methanol extract of the leaves of D. glaucescens furnished a new ecdysteroid 3-deoxy-1 ,20-dihydroxyecdysone (5) [16] together with makisterone A (17), dihydrorubrosterone (18), epi-pterosterone (19) [17] and four oleanane glycosides 3-O-[-D-glucopyranosyl-(1 3)- -D-glucopyranosyl]oleanolic acid 28-O- -D-glucopyranosyl ester (20), 3-O-[-D-xylopyranosyl-(1 2)- -D-glucopyranosyl]oleanolic acid 28-O- -Dglucopyranosyl ester (21) [18], 3-O-{ -D-glucopyranosyl-(1 2)-[-D-glucopyranosyl-(1 3)]- -D-glucopyranosyl oleanolic acid 28-O- -D-glucopyranosyl ester (22) and 3-O-{ -D-glucopyranosyl-(1 3)-[-D-xylopyranosyl-(1 2)]- -D-glucopyranosyl} oleanolic acid 28-O- -D-glucopyranosyl ester (23) [17]. The new ecdysteroid (5) showed 40% potency of 20-hydroxyecdysone in the spiracle index assay using the fourth instar larvae of the silkworm Bombyx mori [16]. Chromatographic separation of the non-quaternary alkaloidal fraction of the methanol extract of D. glaucescens furnished a novel pyridine ring-containing ecdysteroid, named diploclidine (24) [19].

R ₂ R ₃		HO OH	$\overset{R_5}{\longleftarrow}_{R_6}$	HOHO		он н
_	R_1	\mathbf{R}_2	R_3	R_4	R ₅	R_6
<u>1</u>	Н	OH	OH	Н	Н	OH
<u>4</u>	Н	Н	OH	OH	Н	OH
<u>5</u>	OH	OH	Н	Н	Н	OH
<u>6</u>	Н	Н	OH	Н	Н	OH
<u>7</u>	Н	OH	OH	Н	C_2H_5	OH
<u>14</u>	Н	OH	glc-O	Н	Н	OH
<u>17</u>	Н	OH	OH	Н	CH_3	OH
<u>19</u>	Н	OH	OH	Н	OH	Н



Pometia eximia Hook f. of the family Sapindaceae is a tree of moderate size growing in Sri Lanka. Preliminary studies of the methanol extract of the stem of the plant showed strong molluscicidal and larvicidal activities. At a minimum concentration of 15 ppm, the methanol extract caused 100% mortality of Biomphalaria glabrata snails, one of the intermediate hosts of Schistosoma parasite. At a 300 ppm, the methanol extract caused 84% mortality of Aedes albopictus larvae within 24 hours. No antileukaemic activity was observed against L-1210 cells in vitro. The methanol extract also showed highly positive froth test indicating the presence of saponins. Chromatographic separation of the methanol extract furnished hederagenin (25) and nine hederagenin glycosides: 3-O- -L-arabinopyranosyl (26), 3-O- -D-xylopyranosyl-(1 3)- -L-arabinopyranosyl (27), 28-O- -D-apiosyl-(1 2)- -D-glucopyranosyl (28), 3-O- -L-arabinofuranosyl-(1 3)-[-L-rhamnopyranosyl-(1 2)]- -D-xylopyranosyl (29), 3-O- -D-apiosyl-(1 3)-[-L-rhamnopyranosyl-(1 2)]- -D-glucopyranosyl (30), 3-O- -L-arabinofuranosyl-(1 3)-[-L-rhamnopyranosyl-(1 3)-[-L-rhamnopyranL-rhamnopyranosyl-(1 2)]- -L-arabinopyranosyl (31), 3-O- -D-xylopyranosyl-(1 3)-[-L-rhamnopyranosyl-(1 2)]- -L-arabinopyranosyl (32), 3-O- -D-xylopyranosyl-(1 3)-[-L-rhamnopyranosyl-(1 2)]- -D-glucopyranosyl (33), 3-O- -D-galactopyranosyl-(1 3)- $\begin{bmatrix} -L-rhamno pyranosyl-(1 2) \end{bmatrix}$ - D-glucopyranosyl (34). Saponins 28 – 34 are new natural products [20]. The saponins 26, 27, 29, 31 and 32 which contain arabinose showed strong molluscicidal activity at a concentration of 40 ppm, 40 ppm, 10 ppm, 2.5 ppm and 5 ppm, respectively, whereas the saponins 28, 30, 33 and 34 which contain glucose did not show any molluscicidal activity [21]. However, the presence of arabinose moieties and the absence of the glucose does not seems to be a prerequisite for molluscicidal activity, since certain saponins of D. glaucescens containing glucose showed activity against the same snails B. glabrata [10]. The saponin **31** showed strong insecticidal activity against the brown rice planthopper Nilaparvata lugens [22].



	R ₁	\mathbf{R}_2	\mathbf{R}_3	\mathbf{R}_4
2	Н	CH ₃	Н	COOCH ₃
3	Н	CH ₂ OH	Н	COOCH ₃
8	glc	CH ₂ OH	glc	COOCH ₃
9	glc- ³ glc-	CH ₂ OH	glc	COOCH ₃
10	rha- ² glc- ² glc-	CH ₂ OH	glc	COOCH ₃
11	rha- ² glc- ² glc-	CH ₃	glc	COOCH ₃
12	glc- ² glc-	CH ₃	glc	COOCH ₃
13	xyl- ² glc- ² glc-	CH ₃	glc	COOCH ₃
20	glc- ³ glc-	CH ₃	glc	CH ₃
21	xyl- ² glc-	CH ₃	glc	CH ₃
22	glc- ³ glc ₂ -	CH ₃	glc	CH ₃
	glc			
23	glc- ³ glc ₂ -	CH ₃	glc	CH ₃
			-	
	xyl			
25	Н	CH ₂ OH	Н	CH_3
26	ara-	CH ₂ OH	Н	CH_3
27	xyl- ³ ara-	CH ₂ OH	Н	CH_3
28	Н	CH ₂ OH	api- ² glc-	CH ₃
29	rha- ² xyl ₃ -	CH ₂ OH	Н	CH ₃
	ara(f)			
30	rha- ² glc ₃ -	CH ₂ OH	Н	CH_3
	l api			
31	rha- ² ara ₃ *-	CH ₂ OH	Н	CH ₃
	ara(f)			

Table Contd				
32	rha- ² ara ₃ - xyl	CH ₂ OH	н	CH ₃
33	rha- ² glc ₃ -	CH ₂ OH	Н	CH ₃
34	rha- ² glc ₃ -	CH ₂ OH	Н	CH ₃
api = -D-apiofuranosyl ara* = -L-arabinopyranosyl gal = -D-galactopyranosyl	ara = -L-arabinopyranosyl ara(f) = -L-arabinofuranosyl glc = -D-glucopyranosyl			

gal = -D-galactopyranosyl glc = -D-glucopyranosyl rha = -L-rhamnopyranosyl xyl = -D-xylopyranosyl

Filicium decipiens (Wight et Arn.) Thw. of the family Sapindaceae is a tree of moderate size growing in wet and intermediate zones of Sri Lanka. Preliminary investigations of the dichloromethane, methanol and n-butanol fraction of the methanol extract from the leaves and the stem showed a variety of biological activities, *e.g.*, antifungal, antibacterial and molluscicidal activities [23]. Chromatographic separation of the dichloromethane extract of the stem furnished a new natural product 24-norneohopa-4(23),22(29)-diene-3 ,6 ,7 -triol 7-caffeate (**35**) [24] and the *n*-butanol extract from the methanol extract of the leaves furnished sitosterol -D-glucoside (**36**), 3-*O* - D-glucopyranosylkaempferol (**37**), 3-*O* - D-glucopyranosylquercetin (**38**) and 3-*O*-[-L-rhamnopyranosyl-(1 2)- -D-glucopyranosyl] kaempferol (**39**) [25].

Bridelia retusa (L.) Spreng. of the family Euphorbiaceae is a tree of moderate size growing in Sri Lanka. Roots and stem bark of this plant used in the indigenous system of medicine for the treatment of rheumatism and as an astringent [26]. Antifungal activity guided fractionation of dichloromethane, ethyl acetate and methanol extracts of the stem bark of *B. retusa* furnished new bisabolane sesquiterpenes, (*E*)-4-(1,5-dimethyl-3-oxo-1-hexenyl)benzoic acid (**40**), (*E*)-4-(1,5-dimethyl-3-oxo-1,4-hexadienyl)benzoic acid (**41**), (*R*)-4-(1,5-dimethyl-3-oxo-4-hexenyl)benzoic acid (**42**) and (-)-isochaminic acid (**44**), together with the known (*R*)-4-(1,5-dimethyl-3-oxohexyl)benzoic acid (ar-todomatuic acid) (**43**), 5-allyl-1,2,3-trimethoxybenzene (elemicin) (**45**), (+)-sesamin (**46**) and 4-isopropyl-benzoic acid (cumic acid) (**47**) [27]. Antifungal bioassay on TLC bioautography method [28] revealed that the minimum amount of compounds needed to inhibit the growth of *Cladosporioides* were **40** (50 µg/spot), **41** (25 µg/spot), **42** (5 µg/spot), **43** (25 µg/spot), **44** (10 µg/spot), **45** (No activity), **46** (25 µg/spot), **47** (10 µg/spot). Compound **42** showed the most potent antifungal activity [27].

Artocarpus nobilis Thw. is an endemic tree of the family Moraceae growing in mid country regions of Sri Lanka. This is the only endemic species of the genus Artocarpus in Sri Lanka. Several pyranodihydrobenzoxanthones, chromenoflavonoids, triterpenes have been reported from the bark of the plant [29]. Antifungal activity-guided fractionation of the *n*-butanol extract from the methanol extract of the stem bark of *A. nobilis* with a combination of chromatographic separation furnished two stilbene derivatives, (*E*)-4-isopentenyl-3,5,2',4'-tetrahydroxystilbene (**48**) and (*E*)-4-(3-methyl-*E*-but-1-enyl)3,5,2',4'-tetrahydroxystilbene (**48**) and (*E*)-4-(3-methyl-*E*-but-1-enyl)3,5,2',4'-tetrahydroxystilbene (**49**). Both compounds showed strong antifungal activity at 10 μ g/spot against *C. cladosporioides* when assayed by TLC bio-autography method. Antioxidant properties of **48** and **49** were evaluated against the DPPH radical by TLC bio-autography method, in which both compounds were active at 1 μ g/spot [30].





Antifungal activity guided fractionation of the *n*-butanol extract from the methanol extract of the leaves of *A. nobilis* furnished 2',4',4-trihydroxy-3'-geranylchalcone (**50**), 2',4,4'-trihydroxy-3'-[6-hydroxy-3,7-dimethyl-2(*E*),7-octadienyl]chalcone (**51**), 2',4', 4-trihydroxy-3'-[2-hydroxy-7-methyl-3-methylene-6-octaenyl]chalcone (**52**), 2',3,4,4'-tetrahydroxy-3'-geranylchalcone (**53**), 2',3,4,4'-tetrahydroxy-3'-[6-hydroxy-3,7-dimethyl-2(*E*),7-octadienyl]chalcone (**54**). The chalcones **52** and **54** are new natural products, whereas **50** and **51** are reported for the first time from the family Moraceae. All these compounds **50-54** showed fungicidal activity at **50** (5 µg/spot), **51** (5 µg/spot), **52** (5 µg/spot), **53** (2 µg/spot) and **54** (15 µg/spot) against *C. cladosporioides* on TLC bio-autography method. Compounds **50-54** exhibited strong radical scavenging property (active at 1 µg/spot) towards DPPH radical when assayed by TLC bio-autography method [31].

Chromatographic separation of the *n*-butanol extract from the methanol extract of the root bark of *A. nobilis* furnished four new prenylated flavonoids, artonine E 2'-methyl ether (**58**), isoarotonine E 2'-methylether (**59**), dihydroisoarotonine E 2'-methyl ether (**60**) and artonin V 2'-methyl ether (**61**), together with known artobiloxanthone (**55**), artonine E (**56**) and cycloartobiloxanthone (**57**). All these compounds showed strong radical scavenging properties towards DPPH radical[32].

ACKNOWLEDGEMENTS

We gratefully acknowledge the contributions of the members of our research group Ms. B.M.M. Kumarihamy, Mr. A.G.D. Bandara, Ms. C.P. Jayasooriya, Ms. B.A.I.S. Balasooriya

JAYASINGHE AND FUJIMOTO

(Institute of Fundamental Studies), Mr. N. Hara (Tokyo Institute of Technology) and all others whose names appear in the reference section and the acknowledgement sections of the relevant references.

REFERENCES

- Bandaranayake, W.M.; Sultanbawa, M.U.S.; Weerasekera, S.C.; Balasubramaniam, S.; A glossary of sinhala and tamil names of the plants of Sri Lanka, *The Sri Lanka Forester*, **1974**; XI, 67.
- [2] Abeywickrama, B.A.; Proc. Workshop on Natural Products, Colombo, Sri Lanka. 1975.
- [3] Chopra, R.N.; Nayar, S.L.; Chopra, I.C.; In Glossary of Indian Medicinal Plants, Council of Scientific and Industrial Research, New Delhi. 1956, pp. 72 and 99
- [4] Miller, R. W.; Clardy, J.; Kozlowski, J.; Mikolajczak, K. L.; Plattner, R. D.; Powell, R. G.; Smith, C. R.; Weisleder, D.; Qi-Tai, Z.; Planta Medica, 1985, 51, 40–42.
- [5] Shah, V.C.; D'Sa, A.S.; De Souza, N.J.; Steroids, 1989, 53, 559-565.
- [6] Bandara, B.M.R.; Jayasinghe, L.; Karunaratne, V.; Wannigama, G.P.; Bokel, M.; Kraus, W.; Sotheeswaran, S.; *Phytochemistry*, **1989**, 28, 1073 – 1075.
- [7] Jayasinghe, U.L.B.; Wannigama, G.P.; Balasubramaniam, S.; Nasir, H.; Atta-ur-Rahman; Journal of the National Science Council of Sri Lanka, 1992, 20, 187 - 190.
- [8] Bandara, B.M.R.; Jayasinghe, U.L.B.; Karunaratne, V.; Wannigama, G.P.; Bokel, M.; Sotheeswaran, S.; Planta Medica, 1990, 56, 290-292.
- [9] Bandara, B.M.R.; Jayasinghe, L.; Karunaratne, V.; Wannigama, G.P.; Kraus, W.; Bokel, M.; Sotheeswaran, S.; *Phytochemistry*, **1989**, *28*, 2783-2785.
- [10] Jayasinghe, U.L.B.; Wannigama, G.P.; MacLeod, J.K.; Natural Product Letters, 1998, 2, 249-253.
- Jayasinghe, U.L.B.; Wannigama, G.P.; MacLeod, J. K.; Journal of the Chemical Society of Pakistan, 1998, 20, 131-137.
- [12] Jayasinghe, L.; Kumarihamy, B.M.M.; Arundathie, B.G.S.; Dissanayake, L.; Hara, N.; ; Fujimoto, Y.; Steroids, 2003, 68, 447-450.
- [13] Jayasinghe, L.; Hara, N.; Fujimoto, Y.; Phytochemistry, 2003, 62, 563-567.
- [14] Jayasinghe, U.L.B.; Balasooriya, B.A.I.S.; Fujimoto, Y.; Proceedings, Sri Lanka Association for the Advancement of Science (SLAAS), Colombo, 2003, p.216.
- [15] Jayasinghe, U.L.B.; Fujimoto, Y. **2004**, (Manuscript submitted).
- [16] Jayasinghe, L.; Jayasooriya, C. P.; Oyama, K.; Fujimoto, Y.; Steroids, 2002, 67, 555-558.
- [17] Jayasinghe, U.L.B.; Jayasooriya, C.P.; Fujimoto, Y.; Proceedings, Sri Lanka Association for the Advancement of Science (SLAAS), Colombo, 2003, p.220.
- [18] Jayasinghe, U.L.B.; Jayasooriya, C.P.; Fujimoto, Y.; *Fitoterapia*, 2002, 73, 424-427.
- [19] Jayasinghe, L.; Jayasooriya, C.P.; Hara, N.; Fujimoto, Y.; Tetrahedron Letters, 2003, 44, 8769 8771.
- [20] Jayasinghe, L.; Shimada, H.; Hara, N.; Fujimoto, Y.; *Phytochemistry*, **1995**, *40*, 891 897.
- [21] Jayasinghe, U.L.B.; Fujimoto, Y.; Hostettmann, K.; Natural Product Letters, 1998, 12, 135 138.
- [22] Jayasinghe, U.L.B.; Fujimoto, Y.; *Fitoterapia*, **1999**, *70*, 87 88.
- [23] Jayasinghe, U.L.B.; Bandara, A.G.D.; Proceedings, Sri Lanka Association for the Advancement of Science (SLAAS), Colombo, 1997, p.338.
- [24] Jayasinghe, U.L.B.; Bandara, A.G.D.; Hara, N.; Fujimoto, Y.; *Fitoterapia*, 2001, 72, 737-742.
- [25] Jayasinghe, U.L.B.; Balasooriya, B.A.I.S.; Bandara, A.G.D.; Fujimoto, Y.; Natural Product Research, 2004, (in press).
- [26] Jayaweera, D.M.A.; In Medicinal plants used in Ceylon, Part II. The National Science Council of Sri Lanka, Colombo, 1982.
- [27] Jayasinghe, L.; Kumarihamy, B.M.M.; Jayaratne, K.H.R.N.; Udishani, N.W.M.G.; Bandara, B.M.R.; Hara, N.; Fujimoto, Y.; *Phytochemistry*, **2003**, *62*, 637-641.
- [28] Homans, A.L.; Fuchs, A.; Journal of chromatography, 1970, 51, 327–329.
- [29] Sultanbawa, M.U.S.; Surendrakumar, S.; Phytochemistry, 1989, 28, 599–605.
- [30] Jayasinghe, U.L.B.; Puvanendran, S.; Hara, N.; Fujimoto, Y.; *Natural Product Research*, 2003, (in press).
 [31] Jayasinghe, L.; Balasooriya, B.A.I.S.; Padmini, W.C.; Hara, N.; Fujimoto, Y., *Phytochemistry*, 2003, 65, 1287–1290.
- [32] Jayasinghe, L.; Samarakoon, T.B.; Kumarihamy, B.M.M.; Fujimoto, Y.; 2004 (Manuscript submitted).