

Health Beneficial Properties of Potato and Compounds of Interest

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ABSTRACT

Potatoes have shown promising health promoting properties in human cell culture, experimental animal and human clinical studies including antioxidant, hypocholesterolemic, antiinflammatory, antiobesity, anticancer and antidiabetic effects. Compounds present such as the phenolics, fiber, starch, and proteins as well the compounds considered antinutritional such as glycoalkaloids, lectins and proteinase inhibitors are believed to contribute to the health benefits of potatoes. However, epidemiologic studies exploring the role of potatoes in human health have been inconclusive. Some studies support a protective effect of potato consumption in weight management and diabetes while other studies demonstrate no effect, and a few studies suggest a negative effect. Because there are many biological activities attributed to the compounds present in potato, some of which could be beneficial or detrimental depending on specific circumstances, a long term study investigating the association between potato consumption and diabetes, obesity, cardiovascular disease, and cancer while controlling for fat intake is needed.

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INTRODUCTION

Potato (*Solanum tuberosum* L.) belonging to the family Solanaceae is a nourishing food that is rich in calories and biologically active phytochemicals such as β -carotene, polyphenols, ascorbic acid, tocopherol, α -lipoic acid, selenium and dietary fiber¹. The main nutrient in potato is the storage polysaccharide starch. When consumed in whole food form the energy density of potato is similar to legumes². Potatoes are an inexpensive source of energy and good quality protein³. Historically, this was the typical Irish diet, which provided all the important vitamins and nutrients required to support a better life than any other crop when eaten as the sole article of diet⁴.

Potato production has significantly increased in recent years in many countries, especially in Asia where it has become more important as a food and industrial crop². In terms of production, potato ranks sixth in the world after sugar cane, maize, rice, wheat and milk (http://faostat3.fao.org/browse/rankings/commodities_by_regions/E). Until the early 1990s, most potatoes were grown and consumed in Europe, North America and countries of the Soviet Union. Since then, there has been a dramatic increase in potato production and demand in Asia and Africa, where output rose from less than 26 million tons in the early 1960s to more than 217 million tons in 2013. In 2009, the world production of potatoes exceeded 334 million tons and according to the FAO data, the

world production has increased up to approximately 376 million tons in 2013 (http://faostat3.fao.org/browse/rankings/commodities_by_regions/E). Asia and Europe are the world's major potato producing regions, accounting for more than 80 percent of world production in 2013 (http://faostat3.fao.org/browse/rankings/commodities_by_regions/E). The per capita consumption of potato is very high in European countries (87.8 kg/year). Though Asia is the largest producer in the world, the per capita consumption of potato was only 23.9 kg/year in 2005 (<http://www.fao.org/potato-2008/en/world/index.html>). Potato consumption in Asia is on the increase and the demand for potato in Asia is expected to double or triple over the next few years². Although in Asia people are traditionally dependent upon cereals and are generally unaware of the nutritional value of potatoes, in many countries, potatoes are a very significant part of the diet and can make a significant contribution to human nutrition.

In addition to the starch content, potato tubers are rich in proteins, carbohydrates, minerals, vitamins and especially bioactive compounds that contribute to the health beneficial properties of potato². Phytochemicals in potato are concentrated in its peel, and its content is higher in cultivars with brighter peel colors⁵. Phytochemicals play an important role in human health as antioxidants and the high daily consumption of potato contributes to a high phenolic content in our diet⁶. Since the fat content of potato is low, consumption of potato instead of other high carbohydrate containing foods such as rice and pasta may potentially benefit our overall health⁷. Potato has been suggested as a potential functional food by several authors due to presence of several antioxidant compounds in abundant quantities^{1,2,8}.

On the contrary, potatoes are reported to be unhealthy and are often maligned in nutrition because of their suspected link to obesity⁷. This is the reason why their Economic Research Service (ERS) data for per capita availability (PCA) of vegetables, a proxy vegetable consumption, show that vegetable consumption, including consumption of white potatoes, has declined in the past decade⁹. Though the fat content and energy density of potato is similar to legumes, potato is thought to contain more calories and fat compared to rice². Some food guides do not include potatoes in the vegetable group because of their association with high fat diets⁷. To change this negative trend, it is important to point out the nutritionally important components of potato tubers including bioactive compounds, such as polyphenols and carotenoids. Consumption of potato in the right amount with low fat could potentially lead to the prevention of oxidation linked chronic diseases such as type II diabetes and cardiovascular diseases as opposed to its usual form of French fries and potato chips². Therefore, this review will mainly focus on discussing the functional properties of potatoes and their link in preventing the development of chronic diseases.

POTATO AND HEALTH: COMPOUNDS OF INTEREST

Several authors have reviewed the health benefits of potatoes^{2,8,10}. Potatoes which were assumed to play an important role in the development of many chronic diseases in the western world were found to exert many health promoting properties. The beneficial properties exerted by the tubers are reported to be due to the presence of compounds such as phenolics, anthocyanins, resistant starch, dietary fiber, potato proteins and etc². Compounds such as glycoalkaloids and lectins which were considered as anti-nutritional

compounds were also found to exert some health promoting properties^{11,12}. So this part of the review will briefly discuss about some of the compounds which are reported to contribute to the health promoting properties of potato.

Phenolic compounds

Phenolic acids and polyphenols play an important role in human health. Though potatoes are not considered as a food item with high antioxidant activity, the tubers actually present a very significant source of antioxidants in our daily diet due to its high daily consumption¹. Potatoes are reported to be the third most important source of phenols after apples and oranges¹³. Phenolic acids and flavonoids are the most prominent phytochemical groups present in the potato¹⁰. Chlorogenic acid and caffeic acid are the two main phenolic acids present in potato followed by protocatechuic acid, *trans*-cinnamic acid, *para*-coumaric acid, ferulic acid, vanillic acid, gallic acid, syringic acid, and salicylic acid¹⁴. Most of the phenolic acids in potato are present between the cortex and the peel of the potato tuber, and their content reduces towards the center of the tuber¹⁵.

Chlorogenic acid is the predominant phenolic acid (>90%) found in potatoes ranging from 3 to 90 mg/100 g fresh weight (FW) in flesh and 100 to 400 mg/100 g FW in peel¹⁰. Chlorogenic acid is reported to play an important role against the development of diabetes¹⁶, hypertension¹⁷ and also has been reported to exhibit several desirable anticarcinogenic properties^{18,19}. Chlorogenic acid has strong antioxidant activity and

potatoes are an excellent source of this. This partially explains the protective role of potatoes against the development of many chronic diseases.

After chlorogenic acid, caffeic acid is the second most phenolic acid present in potato ranging from 310 to 420 μg per 100 g FW¹⁴. There is a huge variation in the chlorogenic acid content of potatoes based on variety. The chlorogenic acid and caffeic acid content in pigmented cultivars is greater than that in non pigmented cultivars. Stushnoff et al.²⁰ in his study, found approximately 10 fold difference in the chlorogenic acid content of pigmented cultivars and non pigmented cultivars and a 100 fold difference was found for caffeic acid²¹. The cultivars Vitelotte and Luminella were reported to have the highest polyphenol contents (5202 and 572 $\mu\text{g/g}$ dry weight (DW) in the outer flesh, whereas Charlotte and Bintje had the lowest contents (19.5 and 48.0 $\mu\text{g/g}$ DW)²². The yellow PORO3PG6-3 and purple PORO4PG82-1 cultivars had the highest concentration of total phenolics which were 2-fold greater than in the white cultivar Russet Burbank²³. However, in contrast, in a study done by Al-Saikhan et al.²⁴, total phenolics were found to be dependent on genotype and not on flesh color.

Anthocyanins

Anthocyanins belonging to the flavonoids group are present in high amounts in pigmented potatoes¹. Anthocyanins play an important role in human health and are reported to show antioxidant^{25,26}, anticancer²⁷ and antiinflammatory properties²⁸. Anthocyanin content in potatoes has been reported to range between 5.5 and 35 mg/100 g FW². Generally, purple and red fleshed cultivars contain higher amounts of polyphenols than cultivars with a cream or white flesh^{23,29}. The total anthocyanin content in red fleshed potato ranged from 6.9 to 35 mg/ 100 g FW and 5.5 to 17.1 in the

purple fleshed potato¹. Andre et al.³⁰ observed an eleven fold variation in the total phenolic content in Andean potato landraces and the dark purple-fleshed tubers from the Andean cultivar 704429 contained exceptionally high levels of total anthocyanins (16.33 mg g⁻¹ of DW). In a study done by Hamouz et al.²⁹, the purple fleshed varieties Blaue Elise, Blaue St. Galler, Violette, Vitelotte and the red fleshed varieties, Herbie 26, Highland B. Red, Rosalinde, and Rote Emma were found to contain high amount of anthocyanins in the range of 135.3-573.5 mg cyanidin/kg FW. Anthocyanin concentration extending up to 368 mg per 100 g FW was found in the purple-fleshed cv. Urenika³¹.

More than 98% of the total anthocyanins in potatoes are in the acylated form and many are acylated with *para*-coumaric and ferulic acid¹. In colored potatoes, 3-rutinoside-5-glucoside and 3-rutinoside derivatives of pelargonidin, petunidin, malvidin, cyanidin, peonidin and delphinidin have been reported^{31,32,33}. Red fleshed potatoes contain pelargonidin- and peonidin-3-rutinoside-5-glycosides while purple fleshed potatoes are rich in malvidin- and petunidin-3-rutinoside-5-glycosides acylated with *para*-coumaric and ferulic acid^{32,33}. Eichhorn and Winterhalter³⁴ studied the major anthocyanins in four pigmented potato cultivars and pelargonidin was found to be the only anthocyanidin in cv. Highland Burgundy red, malvidin was the predominant aglycon of the cv. Violette and peonidin derivatives were found only in cv. Shetland Black in minor amounts. Except Highland Burgundy Red, Petunidin derivatives were detected in all varieties.

Potato starch

Starch is the major storage polysaccharide in potato. Potatoes contain amylose and amylopectin in a ratio of 1:3⁸. Amylose content of potato cultivars is approximately 31.2% while the content in wild species is 29.7% of the total starch². Amylose is more resistant to digestion than amylopectin. Due to the high amylose content, potato starch is generally resistant to the action of amylase and other amylolytic enzymes in the digestive tract, behaving as a resistant starch⁸. Raw potato starch was shown to be highly resistant to hydrolysis with pancreatic amylase *in vitro*³⁵. Low digestibility of potato starches are due to the granule crystallinity, smaller surface-to- volume ratio of the granules, and the presence of a layer of non-starch barrier material such as polysaccharides on the surface of starch granules³⁶. However, cooking leads to a loss of crystallinity thereby reducing the resistance to amylase digestion². Cooling, freezing and drying make the starch partially resistant to amylase.

In addition to this, phosphorylated potato starches also add to the digestive resistance². Potato starches are highly phosphorylated than other cereal starches³⁷. Potato starch was found to contain 0.2–0.4% (w/w) of monoesterified phosphate groups³⁸. The phosphorus in potato starch is present primarily as phosphate esterified to the glucose residues of the starch³⁹. Phosphate is bound only in the amylopectin fraction of the starch and the glucosyl residues of amylopectin are phosphorylated at either the C6 or C3 position. Most of phosphorylation occurs at the C6 position (70–80% of the phosphorylation), while only 20-30% of phosphorylation occurs at C3 position³⁹. On average, one out of every 200-300 glucose units is reported to be phosphorylated⁴⁰. Noda et al.⁴¹ studied the phosphorous content in 69 potato cultivars and reported a range between 434 to 1087 ppm. The lowest content was observed in Setoyutaka while the highest was in Kachikei No. 11. In his study, the purple- (Inca Purple, Kitamurasaki and

Hokkai No. 92) and red-flesh (Inca Red and Hokkai No. 91) potato cultivars were found to contain relatively higher level of phosphorous (891–1065 ppm). Resistant starch and phosphorylated starch in potatoes have been reported to contribute to blood glucose lowering and cholesterol lowering properties^{37,42}. Enzymatically solubilized polysaccharides from potato pulp have been reported to act as dietary fibers and prebiotics *in vitro*⁴³ and *in vivo*⁴⁴.

Glycoalkaloids

Glycoalkaloids are nitrogen containing steroidal glycosides. The primary glycoalkaloids in potato are α -solanine and α -chaconine, which make up 95% of total glycoalkaloids¹². The other glycoalkaloids found in cultivated potatoes are β - and γ -solanines and chaconines, α - and β -solanines, demissidine, and 5- β -solanidan-3- α -ol, and in wild potatoes leptines, commersonine, demissine, and tomatine³. At certain levels these compounds may be toxic to bacteria, fungi, viruses, insects, animals, and even humans. This toxic effect occurs only when glycoalkaloid intake is very high⁴⁵. The maximum established level for potato glycoalkaloid is 20 mg/100 g FW. However, the glycoalkaloid content of the majority of potato cultivars is between 3 and 10 mg per 100 g of tubers⁴⁵. In addition to their known toxic effects, studies during the past 10 years suggest that they may also possess beneficial effects, including anticancer^{46,47}, antimalarial, antiinflammatory effects and antiglycemic effects² depending on the dose and conditions of use.

Potato Fiber

Dietary fiber plays an important role in human health by acting as a bulking agent and increasing the intestinal mobility and hydration of the feces, binding of unwanted materials such as carcinogenic and a mutagenic substance, facilitate digestion, act as a growth medium for beneficial intestinal microflora, and is reported to exert hypoglycemic, hypocholesterolemic and anticancer effect⁴⁸⁻⁵¹. The European Prospective Investigation on Cancer study reported the protective effect exerted by fiber against colorectal cancer and other health problems was irrelevant of the source of fibre⁵².

Dietary fiber in potatoes is made of cellulose, hemicelluloses, pectins and lignin and other substances which are resistant to digestive enzymes⁵³. In the UK, next to cereals and vegetables, potatoes are the main contributor of dietary fiber in all age groups of men and women⁵². Dietary fiber makes up approximately 2.5% fresh mass of the tuber and is concentrated in the peel where approximately 50% of potato peel is dietary fiber⁴⁸. According to Buttriss and Stokes⁵², the non-starch polysaccharide content or else, the dietary fiber content of baked potato with and without skin was 1.4 and 2.7 g/100g FW respectively.

Cooking, cooling and storage of potatoes are reported to produce retrograded starch⁵². Microwave-heating and deep-fat frying of potatoes was found to reduce the amount of *in vitro* digestible starch and significantly increase both the resistant starch and water-insoluble dietary fiber while the soluble dietary fiber content was unchanged⁵⁴. Soluble dietary fibers are reported to decrease the rate of gastric emptying and intestinal transit time thereby, slowing down the rate of digestion and glucose absorption by the intestine through forming viscous solutions³⁶. Potato is a good source of the water soluble dietary fiber, pectin, which is almost completely metabolized by colonic bacteria⁵⁵. Baking and

extrusion cooking increased the non-starch polysaccharides in potato peels while the soluble to insoluble dietary fiber ratio was increased in the latter⁵⁵. Varo et al.⁵⁶ reported that heat treated potato samples contained more water insoluble dietary fiber and less starch than raw samples.

The addition of viscous dietary fiber to a carbohydrate meal reduces the glycemic response of a meal⁵⁷. Lightowler and Henry⁵⁷, investigated mashed potatoes containing 1, 2 or 3% levels of high-viscosity hydroxypropylmethylcellulose, a modified cellulose dietary fiber and observed significant reduction in glycemic responses in all samples than the standard mashed potato. Dietary fiber preparations from potato skin and flesh were tested for their ability to adsorb the hydrophobic mutagen, 1,8-dinitropyrene (DNP) *in vitro*. Potato skin walls were found to strongly adsorb DNP whereas, in contrast, the flesh walls of potato adsorbed only a small proportion of the DNP and a large increase in the proportion of DNP was found in solution⁵⁸.

Potato protein and peptides

The major groups of potato proteins present in potato tuber are patatins, protease inhibitors, and other proteins⁵⁹. Patatin has been shown to possess antioxidant activity and also inhibit hydroxyl radical induced DNA damage *in vitro*^{24,60-62}. Since peptides have lower molecular weight and less complex structures compared to proteins, their solubility, digestibility and absorbability are higher than those of proteins⁶³. Peptides isolated from potato protein have been reported to exhibit antioxidant⁶³, anticancer⁶⁴, antiobesity⁶⁵, antihyperlipidemic⁶⁶, antifungal/antibiotic activity⁶⁷ and was also reported to exhibit angiotensin-converting enzyme (ACE) inhibition *in vitro*⁶⁸.

Potato protease inhibitors

Protease inhibitors (PI) represent approximately 50% of the total amount of proteins in potato juice⁶⁹. The most studied protease inhibitors from potato tuber are potato protease inhibitor I (PI-1), potato protease inhibitor II (PI-2) and potato carboxypeptidase inhibitor (PCI)⁶⁹. Although protease inhibitors were considered as an antinutritional compound, they have regained interest in recent years because of their exerted anticancer⁶⁴ and antiobesity activity⁷⁰. PI's have been reported to show anticancer activity by preventing tumor cell proliferation, H₂O₂ formation and by protecting from the effect of solar UV irradiation⁶⁹. It acts as a satiety agent by enhancing the release of cholecystokinin⁷¹.

METABOLIC EFFECTS

Potatoes have received much interest recently owing to their observed biological effects *in vitro* such as free radical scavenging^{72,73}, modulation of enzymatic activity⁷⁴, and inhibition of cellular proliferation^{18,51,75}. There is an extensive literature describing each of these biological properties (Table 1). We have focused on the biological effects of potatoes, including their antioxidant, antiobesity, antidiabetic, anticancer, antiinflammatory, antihyperlipidemic and antihypertensive activities, which are commonly ascribed to help explain their potential role against the development of non

communicable diseases such as cardiovascular disease, type 2 diabetes mellitus (T2D), cancer and etc.

Antioxidant activity

Health promoting properties such as antidiabetic, antithrombotic, anticancer, antimutagenic, antiinflammatory and antiallergic activities of plant extracts result from powerful antioxidant and free radical scavenging properties of phenolic compounds^{72-73,76}. Potato tubers are one of the richest sources of antioxidants in the human diet. For example, in the USA, among all the fruits and vegetables consumed potatoes insure an average daily intake of about 64 mg polyphenols per capita¹. The main potato antioxidants are polyphenols (1.226–4.405 mg/kg), ascorbic acid (170–990 mg/kg), carotenoids (as high as 4 mg/kg), tocopherols (0.5–2.8 mg/kg), selenium (0.01 mg/kg) and α -lipoic acid¹. Chlorogenic acid is the primary phenolic compound (being more than 90% of phenolics) found in potatoes^{10,22,77}. In a US study, total phenolic content of peeled potato was 28 mg/100 g FW and it was ranked twentieth out of 23 commonly consumed vegetables and was ranked ninth in terms of antioxidant activity⁷⁸. Flavonoids and flavones extracts from potatoes show high scavenging activities toward oxygen radicals⁷⁹.

Potato chlorogenic acid has been found to be an effective inhibitor of lipid oxidation²⁴. Antioxidant capacity has been directly related to anthocyanin content in potatoes²⁵. Hypercholesterolemic rats fed 300 g of purple potato flakes experienced significantly lower thiobarbituric acid reactive substance (TBARS) levels in serum and liver, and

antioxidant enzyme activities in the liver than those in the control and white potato groups⁸⁰. The serum urate levels in all the flake groups were significantly lower than that in the control group. An extract from purple potato (Hokkai no. 91) ameliorated galactosamine-induced liver damage in rats⁸¹ and was suggested to show hepatoprotective effects via inhibition of lipid peroxidation and/or inflammation in rats. Red potato flakes improved the antioxidant system by enhancing hepatic superoxide dismutase mRNA in rats⁸².

Potato peel extracts (PPE) possess strong antioxidant activity. Almost 50% of phenolics are located in the peel and adjoining tissues and decrease toward the center of the tuber¹⁵. Probably due to the strong antioxidant activity of the anthocyanins, extracts prepared from red peels have stronger activity than those from brown peels⁸³. Potato peel extract inhibited lipid peroxidation induced by FeSO₄ and ascorbic acid in rat red blood cells (RBC) and human RBC membranes with similar effectiveness (80-85% inhibition) at a concentration of 2.5 mg/ml⁷³. The scanning electron microscopy results demonstrated that PPE significantly protected rat RBC against H₂O₂ induced morphological changes and inhibited oxidative damage to human erythrocytes. Additionally, some studies have reported the use of potato peel as a source of antioxidant for the prevention of oxidation of meat products⁸⁴, soy bean oil⁸⁵ and fish oil⁸⁶.

Potato protein hydrolysate has demonstrated potent antioxidant activity *in vitro* and *in vivo*^{63,87}. Intact and hydrolyzed potato proteins lowered the production of peroxide and TBARS values in beef and hydrolyzed potato protein reduced oxidant-induced biochemical changes of pork myofibril protein isolate⁸⁸. Three peptides (5A, 5C and 6C), purified from potato protein hydrolysate fractions, inhibited linoleic acid oxidation

and lipid oxidation in the erythrocyte membrane and oral administration of the peptides reduced ethanol induced gastric mucosal damage in rats⁶³.

Antiobesity activity

From 1991 to 1999, Schulze et al.⁸⁹ examined the association between dietary patterns and weight gain in women enrolled in the Nurse's Health Study II. Western dietary pattern (included red and processed meats, refined grains, sweets and desserts, and potatoes), was associated with weight gain while the prudent dietary pattern (included fruits, vegetables, whole grains, fish, poultry, and salad dressing) facilitated weight maintenance. Mozaffarian et al.⁹⁰ followed up on the Schulze et al.⁸⁹ study and evaluated weight gain in 4 year intervals between 1986 and 2006 in 3 separate prospective cohorts. Participants gained weight with an increase in the intake of potato chips, fried potatoes, sugar-sweetened beverages and unprocessed and processed meats. Anyhow, the 2 prospective cohort studies did not consider the effect of the fat consumed along with potato which can also contribute to weight gain.

Potato proteinase inhibitor II (PI2) reduced food intake in humans when administered orally^{65,70}. At a 1.5 g dose before a meal, PI2 reduced energy intake in healthy subjects⁶⁵, while an average 2 kg weight loss was demonstrated in overweight women when PI2 was taken daily prior to lunch and dinner for four weeks⁹¹. PI2 has been reported to enhance the release of cholecystokinin (CCK) which induces satiety^{71,92}. CCK is a natural signaling peptide released by the gut in response to food. Once

released, CCK acts on various target organs, resulting in signals to the brain, where it induces feeling of fullness and satiety^{65,93}.

Several studies have shown that intact proteins or their hydrolysates could stimulate the secretion of CCK from enteroendocrine cells⁹⁴⁻⁹⁷. A potato extract (Potein) containing 60% carbohydrate and 20% protein including trypsin inhibitory proteins was examined for its effect on food intake and CCK secretion in enteroendocrine cells in rats⁷¹. Oral administration of the extract was found to suppress food intake and stimulate CCK secretion by direct stimulation on enteroendocrine cells and through inhibition of luminal trypsin. In contrast, in another study, feeding of a crude potato PI concentrate was suggested to enhance CCK response primarily by inhibition of trypsin like proteolytic activity in the small intestine and not by direct stimulation of CCK producing cells⁹². The potato extract (Potein) included proteins other than trypsin inhibitors and other non-protein components. Thus presence of some of these components might be responsible for the direct stimulation of CCK secretion. It is likely that CCK secretion is the result of both the direct action of an active compound on CCK producing cells and luminal protease inhibition *in vivo*⁹⁸.

Yoon et al.⁷⁴ studied the antiobesity mechanism of a new purple potato variety in Sprague-Dawley rats. The rats were fed a high fat diet (HFFD) with ethanol extract of *Solanum tuberosum* L. cv. Bora Valley (ESTBV). ESTBV showed potential antiobesity activity via inhibition of lipid metabolism through downregulating the expression of p38 mitogen activated protein kinase (MAPK) and the expression of uncoupled protein 3 (UCP-3). In a study done by Kubow et al.⁹⁹, both sexes of C57BL/6J mice were fed a high fat diet (HFD) for 10 weeks with and without polyphenolic rich potato extracts (PRPE) of cultivars Onaway and Russet Burbank. PRPE attenuated weight gain in male

and female mice by as much as 63.2% and 55.75% respectively. The reduced body weight gain in mice treated with PRPE was mostly due to reduction in adiposity. These results demonstrated greater potency of weight reduction than reported for anthocyanins-rich extracts of the purple fleshed potatoes in HFD fed rats⁷⁴.

Pharmacological treatment of obesity has become widely used in most countries, although the number of available drugs is still very limited. The FDA approved antiobesity drugs, orlistat and sibutramine are widely used and has been reported to show a weight loss of <5 kg in a 1-4 year randomized, placebo controlled trials¹⁰⁰. PI-2 found in potatoes is commercially available for weight loss applications (Eg; Slendesta, SuprxTM, Sola thin). Clinical trials indicate PI-2 is a safe and effective natural agent that promotes satiety and healthy weight loss. When consumed as recommended, potato extract has resulted in statistically significant weight loss and reductions in waist and hip measurements¹⁰¹.

Antidiabetic effect

The glycemic index (GI) of a food depends on the carbohydrate source and amount, the degree of starch gelatinization, and on the type and amount of fiber present⁸. Potato is a carbohydrate rich crop cultivated widely around the world. The glycemic value of potato and potato products vary widely based on the cultivar, maturity, starch structure, processing, and storage conditions³⁶. According to published literature, the GI values of potatoes range from very low, that is, 23 to 41 in unspecified cultivars of potatoes grown in Africa, India and Romania¹⁰² to very high as 144 in boiled Desiree¹⁰³. The GI value of boiled red potatoes (consumed cold), baked US Russet potatoes, instant mashed

potatoes and boiled red potatoes was 56, 77, 88 and 89 respectively¹⁰⁴. Henry et al.¹⁰⁵ examined the GI values of eight varieties of commercially available potatoes in Great Britain and reported a range from 56 to 94. In his study, he found that consumption of baked potatoes along with fat lowered the GI of potatoes (Estima) by 58%, changing the GI classification from high to low GI while GI of potatoes was only lowered by 18% when taken along with protein with classification remaining as high GI. In addition, some wild cultivars grown in Australian Aboriginal regions are reported to exhibit lower GI values than the commercially grown potato cultivars in the Western world³⁶. Based on available studies, potato is the food which yields the most variable glycemic response. However, despite of all these facts, potatoes, as a whole are categorized as a high GI food which requires reconsideration.

In a study done by Montonen et al.¹⁰⁶, potatoes consumed along with butter and whole milk was reported to be associated with increased risk of T2D. Fried potatoes were found to be associated with increased risk of T2D in both men and women¹⁰⁷. Data from the Nurse's Health Study reported a positive association between intake of potatoes and diabetes risk in obese women. It was found that substituting 1 serving of potatoes per day for 1 serving of whole grains increased the risk of T2D by 30%¹⁰⁸.

In contrast, potato intake was found to be associated with a lower risk of T2D in the Shanghai Women's Health Study, where rice was found to increase the risk¹⁰⁹. The contradictory findings of these studies were explained based on a relatively low intake of potato in the Chinese population and the lower amounts of fat added to potatoes during the cooking process by Chinese women. Some other reports also show an inverse relationship between potato intake and risk of T2D¹¹⁰⁻¹¹¹ and some studies also found no association between potato intake and risk of T2D¹¹²⁻¹¹⁴.

To date, most of the studies on potato intake and diabetes focused on a potential positive association. Anyhow, digestibility of raw uncooked potato starch was reported to be poor compared to most cereal starches where, in raw state, 87% of potato starch resists digestion while cereal starches are digested slowly but completely and absorbed *in vivo*¹¹⁵. In addition, potato peel extracts, which are rich in polyphenolic antioxidants were reported to reduce hyperglycemia, oxidative stress, and overall food intake in diabetic rats¹¹⁶. Potatoes are rich in chlorogenic acid and caffeic acid which are implicated in the prevention of T2D¹¹⁷. Chlorogenic acid slows down the release of glucose into the blood stream¹⁶, hence could be helpful in lowering the GI of potatoes. More studies are needed to confirm the link between potato dietary fiber and polyphenolic content in prevention or therapy for T2D and also development of potato cultivars with high resistance to digestion will open up a new avenue for low GI potatoes which would be a good source for diabetic patients and may even decrease the risk of T2D.

Antiinflammatory effect

Markers of systemic inflammation include C-reactive protein (CRP) and the proinflammatory cytokine interleukin-6 (IL-6). Data from subjects (age ≥ 6 years) participating in the 1999-2000 and 2001-2002 National Health and Nutrition Examination Survey (NHANES) reported an association between consumption of whole plant foods including potatoes and lower CRP levels¹¹⁸. The ethanolic extract of potato tubers significantly inhibited both carrageenan- and formalin-induced inflammation in mice as well as arachidonic acid induced ear edema in mice¹¹⁹. In the Busselton Health Study, when BMI was controlled potato intake was associated with lower CRP levels¹²⁰.

In the first human study to investigate the effects of potato consumption on inflammation, Kaspar et al.²⁸ showed that pigmented potatoes were antiinflammatory and lowered plasma concentrations of CRP, 8-hydrodeoxyguanosine, and IL-6 in healthy men compared with men fed with a white potato diet. In a recent study, potato glycoalkaloids and potato peel extracts were found to possess antiinflammatory properties *in vitro*¹²¹.

Antihyperlipidemic effect

There is some evidence that potato protein^{66,122}, resistant and phosphorylated starch^{37,42}, potato fiber^{50,123}, glycoalkaloids¹²⁴ and phenolic compounds¹⁵ contribute to cholesterol lowering properties of potato. Rats fed a potato enriched diet for 3 weeks had lower plasma cholesterol and triglycerides and reduced liver cholesterol than control rats¹²³. In another rat feeding study, Robert et al.¹²⁵ compared the effect of potato, starch, and sucrose-based diets on lipid metabolism. Compared to starch and sucrose diets, consumption of cooked potatoes for 3 weeks lowered cholesterol and triglycerides, and enhanced the antioxidant status and short chain fatty acids in the potato fed animals.

Some studies have reported the positive effect of raw potato starch on plasma and liver lipids^{126,127}. Unlike gelatinized potato starch, raw potato starch is resistant to digestion and acts as dietary fiber in the digestive tract. Retrograded starch from two varieties of potato pulp lowered serum total cholesterol and triglyceride levels compared to controls⁴². Rats fed Benimaru potato showed reduced cholesterol levels and higher levels of fecal bile acids, while rats fed Hokkaikogane potato exhibited reduced triglyceride concentrations and lower hepatic mRNA level of fatty acid synthase and of

sterol regulatory element-binding protein-1c (SREBP1c)⁴². Potato starches are naturally highly phosphorylated, which possibly avoids attack by digestive enzymes and control serum lipids⁴². Gelatinized potato starch containing high level of phosphate was found to reduce serum free fatty acids, triglycerides, and liver triglycerides³⁷. Potato starch increased fecal bile acid excretion but had no effect on cecal short chain fatty acid synthesis or pH. So, the lipid lowering properties was suggested to be due to slow digestion of gelatinized-high-phosphorus-potato starch and not due to the cecal fermentation promoting properties of resistant starch³⁷.

Compared to soy and casein, feeding potato protein to rats resulted in reduced serum total cholesterol, increased fecal bile acid, and neutral steroid excretion¹²⁸. The lipid lowering property was related to the relatively lower methionine content and methionine-to-glycine ratios in potato protein. Lower concentrations of plasma cholesterol and low-density lipoprotein (LDL) were also reported in pigs fed potato protein for 3 weeks¹²⁹. Liyanage et al.¹²² studied the hypocholesterolemic effect of potato peptides. Rats fed a cholesterol free diet containing 20% potato peptides showed greater serum HDL-cholesterol and fecal steroid output and less non-HDL cholesterol. The results were attributed to inhibition of cholesterol absorption, possibly via suppression of micellar solubility of cholesterol. In another study, where hypercholesterolemia was induced in rats, potato peptides reduced the serum non HDL-cholesterol level by stimulating fecal steroid excretion, accelerated by cecal short chain fatty acids⁶⁶.

Whole potato is a rich source of dietary fiber. Camire et al.⁴⁹ evaluated the bile acid binding ability of potato peels *in vitro*. Than the drug cholestyramine, all peels bound a smaller percentage of bile acids. Extrusion cooking of peel enhanced the binding of

cholic, deoxycholic, and glycocholic acids and binding of deoxycholic acid was highly correlated with total dietary fiber and insoluble dietary fiber content. Feeding of potato peel reduced plasma total cholesterol in rats^{50,123}. The authors ascribed the hypolipidemic activity to its fiber content. But it is likely that the polyphenol content and other antioxidants¹⁵ as well as glycoalkaloids contributed to the observed hypocholesterolemia since both tomato and potato glycoalkaloids have a strong affinity for cholesterol¹²⁴.

Anticancer effect

Several studies have shown reduction in proliferation of cancer cells when treated with potato extracts^{18,51,75,130}. Phenolic acids, anthocyanins, fiber, glycoalkaloids and proteinase inhibitors identified in potatoes have been implicated in suppression of cancer cell proliferation *in vitro*^{64,131-133} and *in vivo*^{27,134}. Commercially available potato fiber extract (Potex) was reported to exhibit antiproliferative effect in several tumor cell cultures⁵¹. The fiber extract decreased cancer cell motility, induced apoptosis and also caused morphological changes in tumor cells⁵¹. Colored flesh potatoes are a rich source of anthocyanins with a wide array of health benefits¹. Purple fleshed potatoes were reported to suppress proliferation and elevate apoptosis of colon cancer cells compared with white and yellow fleshed potatoes¹³⁰. Anthocyanins in steamed purple and red potatoes suppressed the growth of benzopyrene-induced stomach cancer in mice¹³⁴.

Extracts from four specialty potatoes and the anthocyanin fraction from genotype CO112F2-2 showed potent antiproliferative properties via increasing the levels of cyclin-dependent kinase inhibitor p27 in both androgen dependent (LNCaP) and

androgen-independent (PC-3) prostate cancer cell lines¹³¹. Anthocyanin fraction of the potato extract induced mitochondrial release and nuclear uptake of the proapoptotic Endo G and AIF proteins. *Solanum jamesii* tuber extracts showed antiproliferative and cytotoxic effects against HT-29 human colon cancer and LNCaP human prostate cancer cell lines¹³³. Red pigmented Mountain Rose cultivar, rich in chlorogenic acid derivatives and anthocyanins showed greater inhibition of carcinogenesis in rats with chemically induced breast cancer as compared with white Russet Burbank cultivar²⁷.

Potato polyphenols are effective against human liver, colon, and prostate cancer cells^{75,133}. Studies with individual phenolics suggested that chlorogenic acid may be the primary compound responsible for the antiproliferative activity. In JB6 mouse epidermal cell line, chlorogenic acid suppressed the proliferation of A549 human lung cancer cell lines and blocked UVB- or TPA-induced transactivation of AP-1 and NF- κ B, which are inflammatory mediators linked to cancer¹⁸. Proliferation of colon cancer cells and liver cancer cells *in vitro* was significantly inhibited by chlorogenic acid⁷⁵.

Several studies have reported that potato glycoalkaloids exhibit an inhibitory effect on the growth of human cancer cell lines such as human colon (HT29), liver (HepG2), cervical (HeLa), lymphoma (U937), and stomach cancer cells¹³⁵⁻¹³⁶. Alpha-chaconine was found to be more effective than alpha-solanine¹³⁶. Alpha-chaconine reduced lung cancer metastasis *in vitro* by suppression of phosphoinositide 3-kinase/Akt/NF-kappaB (PI3K/Akt/NF-kappaB) signaling pathway¹³² and induced apoptosis in HT-29 human colon cancer cells through caspase-3 activation and inhibition of ERK 1/2 phosphorylation¹³⁷. In another study, alpha-chaconine and gallic acid in potato extracts were reported to decrease survival and induce apoptosis in LNCaP and PC3 prostate cancer cells⁴⁷. However, in a recent study, potato glycoalkaloids were reported to show

poor apoptotic activity although the cytotoxic effect was equal to certain cancer drugs¹². In addition to this, other compounds such as potato lectin^{11,138} and potato protease inhibitor 1 and 2 were also reported to show anticancer activity⁶⁴.

Antihypertensive effect

High potassium intake is associated with reduced blood pressure (BP). Potatoes are rich in potassium and are very low in sodium⁸. Vinson et al.¹³⁹ studied the effect of potatoes on blood pressure in 18 overweight, hypertensive adult subjects for 4 weeks in a cross-over design. Consumption of purple potatoes reduced systolic and diastolic BP compared to baseline. Proteins isolated from potatoes and potato products exhibit angiotensin-converting enzyme (ACE) inhibitory action. ACE inhibitors prevent the body from producing angiotensin II, a substance that affects the cardiovascular system by narrowing the blood vessels and releasing hormones that can raise blood pressure¹⁴⁰. Autolysis of protein isolates from the potato tuber tissue was found to enhance ACE-inhibition¹⁴¹. Pihlanto et al.⁶⁸ found hydrolysis of potato proteins to increase the ACE-inhibitory potencies. Potato tuber liquid, a by-product of the potato starch industry was found to be a valuable source of ACE-inhibitory peptides⁵⁹. Results of these studies suggest that potato may be a source for bioactive compounds that benefit cardiovascular health.

SUMMARY

The potato is a nutrient dense food that provides significant amount of nutrients without adding too many calories. In populations where potato is the staple food, potatoes are an

important source of starch, phenolic compounds and dietary fiber (when potato is eaten with the skin). Potatoes possess specific properties that benefit human health in various ways. Potato protein, peptides, protease inhibitors, phenolic compounds, anthocyanins, dietary fiber, resistant starches, and phosphorylated starches have been reported to improve lipid profile, blood glucose level, and blood pressure. *In vitro* and *in vivo* studies also suggest that anthocyanins, glycoalkaloids, and lectins from potatoes are anticancer agents. However, as discussed, the *in vitro* and *in vivo* data have produced conflicting results on the association of potato consumption on obesity and development of diabetes. The data from epidemiological studies regarding these matters are far from convincing. However, to clearly understand the impact of potatoes on human health, a long term study investigating the association between potato consumption and diabetes, obesity, cardiovascular disease, and cancer while controlling for fat intake is needed. More information on the bioavailability of phenolic compounds in potatoes is also needed.

References

1. Lachman J and Hamouz K, Red and purple coloured potatoes as a significant antioxidant source in human nutrition - a review. *Plant Soil Environ* **51(11)**: 477-482 (2005).
2. Camire ME, Kubow S and Donnelly DJ, Potatoes and Human Health. *Crit Rev Food Sci Nutr* **49**: 823-840 (2009).
3. Lachman J, Hamouz K, Orsak M and Pivec V, Potato glycoalkaloids and their significance in plant protection and nutrition. *Rostl Vyroba* **47**: 181-1912 (2001).

4. Nunn N and Qian N, The potato's contribution to population and urbanization: evidence from a historical experiment. *Q J Econ* **126**: 593-650 (2011).
5. Zhang C, Ma Y, Zhao X and Mu J, Influence of copigmentation on stability of anthocyanins from purple potato peel in both liquid state and solid state. *J Agric Food Chem* **57**: 9503-9508 (2009).
6. Xu X, Li W, Lu Z, Beta T and Hydamaka AW, Phenolic content, composition, antioxidant activity, and their changes during domestic cooking of potatoes. *J Agric Food Chem* **57**: 10231-10238 (2009).
7. King JC and Slavin JL, White Potatoes, Human Health, and Dietary Guidance. *Adv Nutr* **4**: 393S-401S (2013).
8. McGill CR, Kurilich AC and Davignon J, The role of potatoes and potato components in cardiometabolic health: A review. *Ann Med* **45**: 467-473 (2013).
9. Storey ML and Anderson PA, Contributions of White Vegetables to Nutrient Intake: NHANES 2009–2010. *Adv Nutr* **4(3)**: 335S-344S (2013).
10. Ezekiel R, Singh N, Sharma S and Kaur A, Beneficial phytochemicals in potato-a review. *Food Res Int* **50**: 487-496 (2013).
11. De Mejia EG and Prisecaru VI, Lectins as bioactive plant proteins: a potential in cancer treatment. *Crit Rev Food Sci Nutr* **45**: 425-445 (2005).
12. Kenny OM, Brunton NP, Rai DK, Collins SG, Jones PW, Maguire AR and O'Brien NM, Cytotoxic and apoptotic potential of potato glycoalkaloids in a number of cancer cell lines. *J Agric Sci Appl* **2(4)**: 184-192 (2013).
13. Chun OK, Kim DO, Smith N, Schroeder D, Han JT and Lee CY, Daily consumption of phenolics and total antioxidant capacity from fruit and vegetables in the American diet. *J Sci Food Agric* **85**: 1715-1724 (2005).

14. Reddivari L, Hale A and Miller J, Determination of phenolic content, composition and their contribution to antioxidant activity in specialty potato selections. *Am J Potato Res* **84(4)**: 275-282 (2007).
15. Friedman M, Chemistry, biochemistry, and dietary role of potato polyphenols- A review. *J Agric Food Chem* **45**: 1523-1540 (1997).
16. Bassoli BK, Cassolla P, Borba-Murad GR, Constantin J, Salgueiro-Pagadigorria CL, Bazotte RB, de Silva RS and de Souza HM, Chlorogenic acid reduces the plasma glucose peak in the oral glucose tolerance test: Effects on hepatic glucose release and glycaemia. *Cell Biochem Funct* **26**: 320-328 (2008).
17. Yamaguchi T, Chikama A, Mori K, Watanabe T, Shioya Y, Katsuragi Y and Tokimitsu I, Hydroxyquinone-free coffee: A double-blind, randomized controlled dose-re response study of blood pressure. *Nutr Metab Cardiovasc Dis* **18(6)**: 408-414 (2007).
18. Feng R, Lu Y, Bowman LL, Qian Y, Castranova V and Ding M, Inhibition of activator protein-1, NF-kappa β , and MAPKs and induction of phase 2 detoxifying enzyme activity by chlorogenic acid. *J Biol Chem* **280(30)**: 27888-27895 (2005).
19. Jin UH, Lee JY, Kang SK, Kim JK, Park WH, Kim JG, Moon SK and Kim CH, A phenolic compound, 5-caffeoylquinic acid (chlorogenic acid), is a new type and strong matrix metalloproteinase-9 inhibitor: Isolation and identification from methanol extract of *Euonymus alatus*. *Life Sci* **77**: 2760-2769 (2005).
20. Stushnoff C, Holm D, Thompson MD, Jiang W, Thompson HJ, Joyce NI and Wilson P, Antioxidant properties of cultivars and selections from the Colorado potato breeding program. *Am J Potato Res* **85(4)**: 267-276 (2008).

21. Navarre DA, Pillai SS, Shakya R and Holden MJ, HPLC profiling of phenolics in diverse potato genotypes. *Food Chem* **127(1)**: 34-41 (2011).
22. Deusser H, Guignard C, Hoffmann L and Evers D, Polyphenol and glycoalkaloid contents in potato cultivars grown in Luxembourg. *Food Chem* **135(4)**: 2814-2824 (2012).
23. Kaspar KL, Park JS, Brown CR, Weller K, Ross CF, Mathison BD and Chew BP, Sensory evaluation of pigmented flesh potatoes (*Solanum tuberosum* L.). *Food Nutr Sci* **4**: 77-81 (2013).
24. Al-Saikhan MS, Howard LR and Miller JC, Antioxidant activity & total phenolics in different genotypes of potato (*Solanum tuberosum* L.). *J Food Sci* **60**: 341-343 (1995).
25. Brown C, Culley D, Yang CP, Durst R and Wrolstad R, Variation of anthocyanin and carotenoid contents and associated antioxidant values in potato breeding lines. *J Am Soc Hortic Sci* **130(2)**: 174-180 (2005).
26. He J and Giusti MM, Anthocyanins: natural colorants with health-promoting properties. *Ann Rev Food Sci Technol* **1**: 163-187 (2010).
27. Thompson MD, Thompson HJ, McGinley JN, Neil ES, Rush DK, Holm DG and Stushnoff C, Functional food characteristics of potato cultivars (*Solanum tuberosum* L.): phytochemical composition and inhibition of 1-methyl-1-nitrosourea induced breast cancer in rats. *J Food Comp Anal* **22(6)**: 571-576 (2009).
28. Kaspar KL, Park JS, Brown CR, Mathison BD and Navarre DA, Pigmented potato consumption alters oxidative stress and inflammatory damage in men. *J Nutr* **141**: 108-111 (2011).

29. Hamouz K, Lachman J, Pazderůl K, Tomášek K, Hejtmánková K and Pivec V, Differences in anthocyanin content and antioxidant activity of potato tubers with different flesh colour. *Plant Soil Environ* **57(10)**: 478–485 (2011).
30. Andre CM, Oufir M, Guignard C, Hoffmann L, Hausman JF, Evers D and Larondelle Y, Antioxidant profiling of native Andean potato tubers (*Solanum tuberosum* L.) reveals cultivars with high levels of beta-carotene, alpha-tocopherol, chlorogenic acid, and petanin. *J Agric Food Chem* **55(26)**: 10839-10849 (2007).
31. Brown CR, Antioxidants in Potato. Antioxidants in potato. *Am J Potato Res* **82**: 163-172 (2005).
32. Lewis CE, Walker JRL, Lancaster JE and Sutton KH, Determination of anthocyanins, flavonoids and phenolic acids in potatoes. I: Coloured cultivars of *Solanum tuberosum* L. *J Sci Food Agric* **77(1)**: 45-57 (1998).
33. Naito K, Umemura Y, Mori M, Sumida T, Okada T, Takamatsu N, Okawa Y, Hayashi K, Saito N and Honda T, Acylated pelargonidin glycosides from a red potato. *Phytochem* **47(1)**: 109-112 (1998).
34. Eichhorn S and Winterhalter P, Anthocyanins from pigmented potato (*Solanum tuberosum* L.) varieties. *Food Res Int* **38(8-9)**: 943-948 (2005).
35. Englyst HN and Cummings JH, Digestion of polysaccharides of potato in the small intestine of man. *Am J Clin Nutr* **45**: 423-431 (1987).
36. Nayak B, Berrios JDJ and Juming Tang, Impact of food processing on the glycemic index (GI) of potato products. *Food Res Int* **56**: 35–46 (2014).

37. Kanazawa T, Atsumi M, Mineo H, Fukushima M, Nishimura N, Noda T and Chiji H, Ingestion of gelatinized potato starch containing a high level of phosphorus decreases serum and liver lipids in rats. *J Oleo Sci* **57**: 335-343 (2008).
38. Blennow A, Engelsen SB, Nielsen TH, Baunsgaard L and Mikkelsen R, Starch phosphorylation: a new front line in starch research. *Trends Plant Sci* **7(10)**: 445-450 (2002).
39. Carpenter MA, Joyce NI, Genet RA, Cooper RD, Murray SR, Noble AD, Butler RC and Timmerman-Vaughan GM, Starch phosphorylation in potato tubers is influenced by allelic variation in the genes encoding glucan water dikinase, starch branching enzymes I and II, and starch synthase III. *Front Plant Sci* **6**: 1-12 (2015).
40. Jacobsen HB, Madsen MH, Christianse J and Nielsen TH, The degree of starch phosphorylation as influenced by phosphate deprivation of potato (*Solanum tuberosum* L.) plants. *Potato Res* **41(2)**: 109-116 (1998).
41. Noda T, Tsuda S, Mori M, Takigawa S, Matsuura-Endo C, Kim SJ, Hashimoto N and Yamauchi H, Determination of the phosphorus content in potato starch using an energy-dispersive X-ray fluorescence method. *Food Chem* **95**: 632-637 (2006).
42. Hashimoto N, Ito Y, Han KH, Shimada K, Sekkikawa M, Topping DL, Bird AR, Noda T, Chiji H and Fukushima M, Potato pulps lowered the serum cholesterol and triglyceride levels in rats. *J Nutr Sci Vitaminol* **52**: 445-450 (2006).
43. Thomassen LV, Vignæs LK, Licht TR, Mikkelsen JD and Meyer AS, Maximal release of highly bifidogenic soluble dietary fibers from industrial potato pulp by minimal enzymatic treatment. *Appl Microbiol Biotechnol* **90(3)**: 873-884 (2011).

44. Lærke HN, Meyer AS, Kaack KV and Larsen T, Soluble fiber extracted from potato pulp is highly fermentable but has no effect on risk markers of diabetes and cardiovascular disease in Goto-Kakizaki rats. *Nutr Res* **27**: 152-160 (2007).
45. Lister CE and Munro J, Nutrition and health qualities of potatoes – a future focus. Crop & Food Research Confidential Report No. 143, New Zealand Institute for Crop & Food Research Limited Private Bag 4704, Christchurch, New Zealand. p 47 (2000).
46. Lu MK, Shih YW, Chang Chien TT, Fang LH, Huang HC and Chen PS, α -Solanine inhibits human melanoma cell migration and invasion by reducing matrix metalloproteinase-2/9 activities. *Biol Pharm Bull* **33**: 1685-1691 (2010).
47. Reddivari L, Vanamala J, Safe SH and Miller JC, The bioactive compounds α -chaconine and gallic acid in potato extracts decrease survival and induce apoptosis in LNCaP and PC3 prostate cancer cells. *Nutr Cancer* **62**: 601-610 (2010).
48. Camire ME, Zhao J, Dougherty MP and Bushway RJ, In vitro binding of benzo[a]pyrene by extruded potato peels. *J Agric Food Chem* **43**: 970-973 (1995).
49. Camire ME, Zhao J and Violette DA, In vitro binding of bile acids by extruded potato peels. *J Agric Food Chem* **41**: 2391-2394 (1993).
50. Lazarov K and Werman MJ, Hypocholesterlaemic effect of potato peels as a dietary fiber source. *J Med Sci Res* **24**: 581-582 (1996).
51. Langner E, Rzeski W, Kaczor J, Kandefer-Szerszeń M and Pierzynowski SG, Tumour cell growth-inhibiting properties of water extract isolated from heated potato fibre (Potex). *JPCCR* **3(1)**: 36-41 (2009).
52. Buttriss JL and Stokes CS, Dietary fibre and health: an overview. *Nutr Bull* **33(3)**: 186–200 (2008).

53. Gumul D, Ziobro R, Noga M and Sabat R, Characterization of five potato cultivars according to their nutritional and pro-health components. *Acta Sci Pol Technol Aliment* **10(1)**: 73-81 (2011).
54. Thed ST and Phillips RD, Changes of dietary fiber and starch composition of processed potato products during domestic cooking. *Food Chem* **52(3)**: 301–304 (1995).
55. Dhingra D, Michael M, Rajput H and Patil RT, Dietary fibre in foods: a review. *J Food Sci Technol* **49(3)**: 255–266 (2012).
56. Varo P, Laine R and Koivistoinen P, Effect of heat treatment on dietary fibre: inter laboratory study. *J Assoc Off Anal Chem* **66(4)**: 933–938 (1983).
57. Lightowler HJ and Henry CJ, Glycemic response of mashed potato containing high-viscosity hydroxypropylmethylcellulose. *Nutr Res* **29(8)**: 551-557 (2009).
58. Harris PJ, Robertson AM, Hollands HJ and Ferguson LR, Adsorption of a hydrophobic mutagen to dietary fibre from the skin and flesh of potato tubers. *Mutat Res-Genet Tox* **260(2)**: 203–213 (1991).
59. Pihlanto A and Mäkinen S, Antihypertensive properties of plant protein derived peptides in Food Peptides in Health and Disease, ed. by Blanca Hernandez-Ledesma and Chia-Chien Hsieh. InTech, pp 145-182 (2013).
60. Liu YW, Han CH, Lee MH, Hsu FL and Hou WC, Patatin, the tuber storage protein of potato (*Solanum tuberosum* L.), exhibits antioxidant activity in vitro. *J Agric Food Chem* **51**: 4389-4393 (2003).
61. Arcan I and Yemenicioglu A, Antioxidant activity of protein extracts from heat-treated or thermally processed chickpeas and white beans. *Food Chem* **103**: 301–312 (2007).

62. Sun Y, Jiang L and Wei D, Partial characterization, in vitro antioxidant and antiproliferative activities of patatin purified from potato fruit juice. *Food Funct* **4(10)**: 1502-1511 (2013).
63. Kudo K, Onoderab S, Takedab Y, Benkebliac N and Shiomi N, Antioxidative activities of some peptides isolated from hydrolyzed potato protein extract. *J Funct Foods* **1(2)**: 170-176 (2009).
64. Huang C, Ma WY, Ryan CA and Dong Z, Proteinase inhibitors I and II from potatoes specifically block UV-induced activator protein-1 activation through a pathway that is independent of extracellular signal-regulated kinases, c-Jun N-terminal kinases, and P38 kinase. *PNAS* **94(22)**: 11957-11962 (1997).
65. Hill AJ, Peikin SR, Ryan CA and Blundell JE, Oral administration of proteinase inhibitor II from potatoes reduces energy intake in man. *Physiol Behav* **48**: 241-246 (1990).
66. Liyanage R, Han KH, Shimada KI, Sekikawa M, Tokuji Y, Ohba K, Sasaki K, Jayawardana BC, Shimizu T and Fukushima M, Potato and soy peptides alter caecal fermentation and reduce serum non-HDL cholesterol in rats fed cholesterol. *Eur J Lipid Sci Technol* **111**: 884-892 (2009).
67. Kim JY, Park CS, Hwang I, Cheong H, Nah JW, Hahm KS and Park Y, Protease inhibitors from plants with antimicrobial activity. *Int J Mol Sci* **10**: 2860-2872 (2009).
68. Pihlanto A, Akkanen S and Korhonen HJ, ACE-inhibitory and antioxidant properties of potato (*Solanum tuberosum*). *Food Chem* **109**: 104-112 (2008).
69. Pouvreau L, Gruppen H, Piersma SR, van den Broek LA, van Koningsveld GA and Voragen AG, Relative abundance and inhibitory distribution of protease

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inhibitors in potato juice from cv. Elkana. *J Agric Food Chem* **49(6)**: 2864-2874 (2001).

70. Schwartz JG, Guan D, Green GM and Phillips WT, Treatment with an oral proteinase inhibitor slows gastric emptying and acutely reduces glucose and insulin levels after a liquid meal in type II diabetic patients. *Diabetes Care* **17(4)**: 255-262 (1994).
71. Nakajima S, Hira T, Tsubata M, Takagaki K and Hara H, Potato extract (potein) suppresses food intake in rats through inhibition of luminal trypsin activity and direct stimulation of cholecystokinin secretion from enteroendocrine cells. *J Agric Food Chem* **59(17)**: 9491-9496 (2011).
72. Singh N and Rajini PS, Free radical scavenging activity of an aqueous extract of potato peel. *Food Chem* **85(4)**: 611-616 (2004).
73. Singh N and Rajini PS, Antioxidant-mediated protective effects of potato peel extract in erythrocytes against oxidative damage. *Chem Biol Interact* **173(2)**: 97-104 (2008).
74. Yoon SS, Rhee YH, Lee HJ, Lee EO, Lee MH, Ahn KS, Lim HT and Kim SH, Uncoupled protein 3 and p38 signal pathways are involved in antiobesity activity of *Solanum tuberosum* L. cv. Bora Valley. *J Ethnopharmacol* **118(3)**: 396-404 (2008).
75. Quanyi W, Qin C, Maolong H, Priya M, Junying S and Qing Y, Inhibitory effect of antioxidant extracts from various potatoes on the proliferation of human colon and liver cancer cells. *Nutr Cancer* **63(7)**: 1044-1052 (2011).

76. Mohdaly AA, Sarhan MA, Smetanska I and Mahmoud A, Antioxidant properties of various solvent extracts of potato peel, sugar beet pulp and sesame cake. *J Agric Food Chem* **90(2)**: 218-226 (2010).
77. Valiñas MA, Lanteri ML, ten Have A and Andreu AB, Chlorogenic Acid Biosynthesis Appears Linked with Suberin Production in Potato Tuber (*Solanum tuberosum*). *J Agric Food Chem* **63(19)**: 4902–4913 (2015).
78. Vinson JA, Hao Y, Su X and Zubik L, Phenol Antioxidant Quantity and Quality in Foods: Vegetables. *J Agric Food Chem* **46(9)**: 3630-3634 (1998).
79. Chu YH, Chang CL and Hsu HF, Flavonoid content of several vegetables and their antioxidant activity. *J Sci Food Agric* **80**: 561-566 (2000).
80. Han KH, Matsumoto A, and Shimada K, Effects of anthocyanin-rich purple potato flakes on antioxidant status in f344 rats fed a cholesterol-rich diet. *Br J Nutr* **98**: 914-921 (2007).
81. Han KH, Hashimoto N, Shimada K, Sekikawa M, Noda T, Yamauchi H, Hashimoto M, Chiji H, Topping DL and Fukushima M, Hepatoprotective effects of purple potato extract against d-galactosamine-induced liver injury in rats. *Biosci Biotechnol Biochem* **70(6)**: 1432-1437 (2006).
82. Han KH, Shimada K, Sekikawa M and Fukushima M, Anthocyanin-rich red potato flakes affect serum lipid peroxidation and hepatic SOD mRNA level in rats. *Biosci Biotechnol Biochem* **71(5)**: 1356-1359 (2007).
83. Al-Weshahy A and Venket Rao A, Isolation and characterization of functional components from peel samples of six potato varieties growing in Ontario. *Food Res Int* **42**: 1062-1066 (2009).

84. Kanatt SR, Chander R, Radhakrishna P and Sharma A, Potato peel extract-a natural antioxidant for retarding lipid peroxidation in radiation processed lamb meat. *J Agric Food Chem* **53**: 1499-1504 (2005).
85. Zia-ur-rehman, Habib F and Shah WH, Utilization of potato peels extract as a natural antioxidant in soy bean oil. *Food Chem* **85**: 215-220 (2004).
86. Habeebullah SFK, Nielsen NS and Jacobsen C, Antioxidant activity of potato peel extracts in a fish-rapeseed oil mixture and in oil-in-water emulsions. *J Am Oil Chem Soc* **87(11)**: 1319-1332 (2010).
87. Kudo K, Matsumoto M, Onodera S, Takeda Y, Ando K and Shiomi N, Antioxidative activity and protective effect against ethanol-induced gastric mucosal damage of a potato protein hydrolysate. *J Nutr Sci Vitaminol* **49**: 451-455 (2003).
88. Wang LL and Xiong YL, Inhibition of oxidant-induced biochemical changes of pork myofibrillar protein by hydrolyzed potato protein. *J Food Sci* **73**: C482-C487 (2008).
89. Schulze MB, Fung TT, Manson JE, Willett WC and Hu FB, Dietary patterns and changes in body weight in women. *Obesity (Silver Spring)* **14(8)**: 1444-1453 (2006).
90. Mozaffarian D, Hao T, Rimm EB, Willett WC and Hu FB, Changes in diet and lifestyle and long-term weight gain in women and men. *N Engl J Med* **364**: 2392-2404 (2011).
91. Spiegel TA, Hubert C and Peiken SR, Effect of a pre-meal beverage containing proteinase inhibitor from potatoes on satiety in dieting overweight women (Abstract). Presented at the North American Association for the Study of Obesity

(NAASO) Annual Meeting, University of Medicine and Dentistry of New Jersey (1999).

92. Komarnytsky S, Cook A and Raskin I, Potato protease inhibitors inhibit food intake and increase circulating cholecystokinin levels by a trypsin-dependent mechanism. *Int J Obesity* **35(2)**: 236-243 (2011).
93. Moran TH and Kinzig KP, Gastrointestinal satiety signals II. Cholecystokinin. *Am J Physiol Gastrointest Liver Physiol* **286(2)**: G183-G188 (2004).
94. Cordier-Bussat M, Bernard C, Haouche S, Roche C, Abello J, Chayvialle JA and Cuber JC, Peptones stimulate cholecystokinin secretion & gene transcription in the intestinal cell line STC-1. *Endocrinol* **138(3)**: 1137-1144 (1997).
95. Némoy-Gaillard E, Bernard C, Abello J, Cordier-Bussat M, Chayvialle JA and Cuber JC, Regulation of cholecystokinin secretion by peptones and peptidomimetic antibiotics in STC-1 cells. *Endocrinol* **139(3)**: 932-938 (1998).
96. Foltz M, Ansems P, Schwarz J, Tasker MC, Loubakos A and Gerhardt CC, Protein hydrolysates induce CCK release from enteroendocrine cells and act as partial agonists of the CCK1 receptor. *J Agric Food Chem* **56(3)**: 837-843 (2008).
97. Hira T, Maekawa T, Asano K and Hara H, Cholecystokinin secretion induced by beta-conglycinin peptone depends on Galphaq-mediated pathways in enteroendocrine cells. *Eur J Nutr* **48(2)**: 124-127 (2009).
98. Chen W, Hira T, Nakajima S, Tomozawa H, Tsubata M, Yamaguchi K and Hara H, Suppressive effect on food intake of a potato extract (Potein®) involving cholecystokinin release in rats. *Bio Biotech Biochem* **76(6)**: 1104-1109 (2012).
99. Kubow S, Hobson L, Iskandar MM, Sabally K, Donnelly DJ and Agellon LB, Extract of Irish potatoes (*Solanum tuberosum* L.) decreases body weight gain and

adiposity and improves glucose control in the mouse model of diet-induced obesity. *Mol Nutr Food Res* **58(11)**: 2235-2238 (2014).

100. Padwal R, Kezouh A, Levine M and Etminan M, Long-term persistence with orlistat and sibutramine in a population-based cohort. *Int J Obesity* **31(10)**: 1567-1570 (2007).
101. Dana S, An open label clinical trial to evaluate a satiety aid for weight loss in overweight to obese, healthy adults (koslow trial, phase 1). Kemin Health White paper (2005).
102. Foster-Powell K, Holt SHA and Brand-Miller JC, International tables of glycemic index and glycemic load values. *Am J Clin Nutr* **76**: 5-56 (2002).
103. Soh NL and Brand-Miller J, The glycemic index of potatoes: The effect of variety, cooking method and maturity. *Eur J Clin Nutr* **53**: 249-254 (1999).
104. Fernandes G, Velangi A and Wolever TM, Glycemic index of potatoes commonly consumed in North America. *J Am Diet Assoc* **105**: 557-562 (2005).
105. Henry CJ, Lightowler HJ, Strik CM and Storey M, Glycaemic index values for commercially available potatoes in Great Britain. *Br J Nutr* **94**: 917-921 (2005).
106. Montonen J, Knekt P, Harkanen T, Jarvinen R, Heliovaara M, Aromaa A and Reunanen A, Dietary patterns and the incidence of type 2 diabetes. *Am J Epidemiol* **161(3)**: 219-227 (2005).
107. Liese AD, Weis KE, Schulz M and Tooz JA, Food intake patterns associated with incident type 2 diabetes: the Insulin Resistance Atherosclerosis Study. *Diabetes Care* **32**: 263-268 (2009).

108. Halton TL, Willett WC, Liu S, Manson JE, Stamfer MJ and Hu FB, Potato and French fry consumption and risk of type 2 diabetes in women. *Am J Clin Nutr* **83**: 284-290 (2006).
109. Villegas R, Liu S, Gao YT, Yang G, Li H, Zheng W and Shu XO, Prospective study of dietary carbohydrates, glycemic index, glycemic load, and incidence of type 2 diabetes mellitus in middle-aged Chinese women. *Arch Intern Med* **167**: 2310-2316 (2007).
110. Panagiotakos DB, Pitsavoa C, Arvaniti F and Stefanadis C, Adherence to the Mediterranean food pattern predicts the prevalence of hypertension, hypercholesteremia, diabetes and obesity, among adults; the accuracy of the MedDietScore. *Prev Med* **44**: 335-340 (2007).
111. Morimoto A, Ohno Y, Tatsumi Y, Mizuno S and Watanabe S, Effects of healthy dietary pattern and other lifestyle factors on incidence of diabetes in a rural Japanese population. *Asia Pac J Clin Nutr* **21**: 601-608 (2012).
112. Williams DE, Wareham NJ, Cox BD, Byrne CD, Hales CN and Day NE, Frequent salad vegetable consumption is associated with a reduction in the risk of diabetes mellitus. *J Clin Epidemiol* **52**: 329-335 (1999).
113. Hodge AM, English DR, O'Dea K and Giles GG, Glycemic index and dietary fiber and the risk of type 2 diabetes. *Diabetes Care* **27**: 2701-2706 (2004).
114. Liu S, Serdula M, Janket SJ, Cook NR, Sesso HD, Willett WC, Manson JE and Buring J, A prospective study of fruit and vegetable intake and the risk of type 2 diabetes in women. *Diabetes Care* **27**: 2993-2996 (2004).

115. Holm J, Lundquist I, Bjoerck I, Eliasson AC and Asp NG, Degree of starch gelatinization, digestion rate of starch in vitro, and metabolic response in rats. *Am J Clin Nutr* **47**: 1010–1016 (1988).
116. Singh N, Kamath V and Rajini PS, Attenuation of hyperglycemia and associated biochemical parameters in STZ-induced diabetic rats by dietary supplementation of potato peel powder. *Clin Chim Acta* **353(1-2)**: 165-175 (2005).
117. Paynter NP, Yeh HC, Voutilainen S, Schmidt MI, Heiss G, Folsom AR, Brancati FL and Kao WH, Coffee and sweetened beverage consumption and the risk of type 2 diabetes mellitus: the atherosclerosis risk in communities study. *Am J Epidemiol* **164**: 1075-1084 (2006).
118. Lipsky LM, Cheon K, Nansel TR and Albert PS, Candidate measures of whole plant food intake are related to biomarkers of nutrition and health in the US population (National Health and Nutrition Examination Survey 1999-2002). *Nutr Res* **32**: 251-259 (2012).
119. Choi E and Koo S, Anti-nociceptive & anti-inflammatory effects of the ethanolic extract of potato (*Solanum tuberosum*). *Food Agric Immunol* **16(1)**: 29-39 (2005).
120. Hickling S, Hung J, Knuiman M, Divitini M and Beilby J, Are the associations between diet and C-reactive protein independent of obesity? *Prev Med* **47(1)**: 71-76 (2008).
121. Kenny OM, McCarthy CM, Brunton NP, Hossain MB, Rai DK, Collins SG, Jones PW, Maguire AR and O'Brien NM, Anti-inflammatory properties of potato glycoalkaloids in stimulated Jurkat and Raw 264.7 mouse macrophages. *Life Sci* **92(13)**: 775-782 (2013).

122. Liyanage R, Han KH, Watanabe S, Shimada K, Sekikawa M, Ohba K, Tokuji Y, Ohnishi M, Shibayama S, Nakamori T and Fukushima M, Potato and soy peptide diets modulate lipid metabolism in rats. *Biosci Biotechnol Biochem* **72**: 943-950 (2008).
123. Robert L, Narcy A, Demigne C, Mazur A and Remesy C, Entire potato consumption improves lipid metabolism and antioxidant status in cholesterol-fed rat. *Eur J Nutr* **45(5)**: 267-274 (2006).
124. Friedman M, Potato glycoalkaloids and metabolites: roles in the plant and in the diet. *J Agric Food Chem* **54**: 8655-8681 (2006).
125. Robert L, Narcy A, Rayssiguier Y, Mazur A and Remesy C, Lipid metabolism and antioxidant status in sucrose vs. potato-fed rats. *JACN* **27(1)**: 109-116 (2008).
126. Younes H, Levrat MA, Demigné C and Rémésy C, Resistant starch is more effective than cholestyramine as a lipid-lowering agent in the rat. *Lipids* **30(9)**: 847-853 (1995).
127. Younes H, Coudray C, Bellanger J, Demigne C, Rayssiguier Y and Remesy C, Effect of two fermentable carbohydrates (inulin and resistant starch) and their combination on calcium and magnesium balance in rats. *Br J Nutr* **86**: 479-485 (2001).
128. Morita T, Oh-hashii A, Takei K, Ikai M, Kasaoka S and Kiriyaama S, Cholesterol-lowering effects of soybean, potato and rice proteins depend on their low methionine contents in rats fed a cholesterol-free purified diet. *J Nutr* **127**: 470-477 (1997).
129. Spielmann J, Kluge H, Stangl GI and Eder K, Hypolipidemic effects of potato protein and fish protein in pigs. *J Anim Physiol Anim Nutr* **93(4)**: 400-409 (2009).

130. Madiwale GP, Reddivari L, Holm DG and Vanamala J, Storage elevates phenolic content and antioxidant activity but suppresses antiproliferative and pro-apoptotic properties of colored-flesh potatoes against human colon cancer cell lines. *J Agric Food Chem* **59(15)**: 8155-8166 (2011).
131. Reddivari L, Vanamala J, Chintharlapalli S, Safe SH and Miller JC, Anthocyanin fraction from potato extracts is cytotoxic to prostate cancer cells through activation of caspase dependent and caspase-independent pathways. *Carcinogenesis* **28(10)**: 2227-2235 (2007).
132. Shih YW, Chen PS, Wu CH, Jeng YF and Wang CJ, α -Chaconine-reduced metastasis involves a PI3K/Akt signaling pathway with downregulation of NF- κ B in human lung adenocarcinoma A549 cells. *J Agric Food Chem* **55**: 11035-11043 (2007).
133. Nzaramba MN, Reddivari L, Bamberg JB and Miller JC, Antiproliferative activity and cytotoxicity of *Solanum jamesii* tuber extracts on human colon and prostate cancer cells in vitro. *J Agric Food Chem* **57(18)**: 8308-8315 (2009).
134. Hayashi K, Hibasami H, Murakami T, Terahara N, Mori M and Tsukui A, Induction of apoptosis in cultured human stomach cancer cells by potato anthocyanins and its inhibitory effects on growth of stomach cancer in mice. *Food Sci Technol Res* **12**: 22-26 (2006).
135. Lee KR, Kozukue N, Han JS, Park JH, Chang E, Baek EJ, Chang JS and Friedman M, Glycoalkaloids and metabolites inhibit the growth of human colon (HT29) and liver (HepG2) cancer cells. *J Agric Food Chem* **52**: 2832-2839 (2004).

136. Friedman M, Lee KR, Kim HJ, Lee IS and Kozukue N, Anticarcinogenic effects of glycoalkaloids from potato against cervical, liver, lymphoma, and stomach cancer cells. *J Agric Food Chem* **53**: 6162-6169 (2005).
137. Yang SA, Paek SH, Kozukue N, Lee KR and Kim JA, α -Chaconine, a potato glycoalkaloid, induces apoptosis of HT-29 human colon cancer cells through caspase-3 activation and inhibition of ERK 1/2 phosphorylation. *Food Chem Toxicol* **44**: 839-846 (2006).
138. Wang H, Ng TB, Ooi VEC and Liu WK, Effects of lectins with different carbohydrate-binding specificities on hepatoma, choriocarcinoma, melanoma and osteosarcoma cell lines. *Int J Biochem Cell Biol* **32(3)**: 365-372 (2000).
139. Vinson JA, Demkosky CA, Navarre DA and Smyda MA, High-antioxidant potatoes: acute in vivo antioxidant source and hypotensive agent in humans after supplementation to hypertensive subjects. *J Agric Food Chem* **60**: 6749-6754 (2012).
140. Sweitzer NK, What is an angiotensin converting enzyme inhibitor? *Circulation* **108**: e16-e18 (2003).
141. Makinen S, Kelloniemi J, Pihlanto A, Makinen K, Korhonen H, Hopia A and Valkonen JPT, Inhibition of angiotensin converting enzyme I caused by autolysis of potato proteins by enzymatic activities confined to different parts of the potato tuber. *J Agric Food Chem* **56(21)**: 9875-9883 (2008).
142. Geng F, He Y, Yang L and Wang Z, A rapid assay for angiotensin-converting enzyme activity using ultra-performance liquid chromatography-mass spectrometry. *Biomed Chromatogr* **24(3)**: 312-317 (2010).

143. Sajilata MG, Singhal RS and Kulkarni PR, Resistant starch – A review. *Compr Rev Food Sci Food Saf* **5**: 1–17 (2006).

Table 1. Functional properties and physiological effects of potato compounds

Active ingredient	Functional properties	Physiological effects	References
Phenolic compounds	Antioxidant	Prevent oxidative damage to DNA and other biomolecules; up-regulate expression of cellular antioxidant enzymes.	2, 24
	Antidiabetic	Reduce gut glucose absorption; increase insulin sensitivity; inhibit hepatic glucose-6-phosphatase; reduce oxidative stress and overall food intake.	2, 16, 99, 116
	Antihypertensive	Reduce systolic and diastolic blood pressure; inhibit angiotensin-converting enzyme (ACE).	8, 142
	Anticancer	Prevent proliferation of cancer cells; up-regulate expression of cellular antioxidant enzymes; block inflammatory mediators linked to cancer; suppress ROS-mediated NF- κ B, AP-1 and MAPK activation.	18, 19, 75, 133
	Antiobesity	Inhibit lipid metabolism through down regulation of expression of p38 mitogen activated protein kinase (MAPK) and uncoupled protein 3 (UCP-3); suppress	74, 99

		adipogenesis.	
Anthocyanins	Antioxidant	Prevent oxidative damage to DNA; prevent lipid oxidation; up-regulate expression of cellular antioxidant enzymes	25, 28, 80, 82
	Anticancer	Suppress proliferation and elevate apoptosis; induce mitochondrial release and nuclear uptake of proapoptotic Endo G and AIF proteins; cytotoxic effect against cancer cell lines.	130, 131, 134
	Anti-inflammatory	Reduce plasma concentrations of CRP, 8-hydrodeoxyguanosine and IL-6.	28
Potato starch (Resistant and phosphorylated starch)	Hypoglycemic	Reduce glycemic response of food	115
	Hypolipidemic	Resist digestion; increase fecal bile acid excretion; inhibit synthesis of fatty acids; increase cecal short chain fatty acids synthesis.	37, 42, 143
Glycoalkaloids	Anticancer	Reduce metastasis and induce apoptosis; inhibit proliferation; reduce matrix metalloproteinase-2 (MMP-2) and MMP-9 activity; cytotoxic to cancer cells.	12, 46, 47, 135, 136
	Anti-inflammatory	Reduce interleukin-2 and	121

		interleukin-8 production; reduce NO production.	
Lectins	Anticancer	Induce apoptosis; limit the synthesis of proteins, DNA, and RNA in cancer cells.	2, 11
Fibre	Hypoglycemic	Reduce glycemic response of food; reduce hypertrophy of liver and kidney; normalize activity of antioxidant enzymes.	116
	Hypocholesterolemic		49, 123
	Anticancer	Bind to bile acid and reduce availability, increase cecal short chain fatty acids synthesis; increase neutral steroid excretion. Prevent tumor cell proliferation; reduce cancer cell motility; induce apoptosis; cause morphological changes in tumor cells; adsorb mutagens.	50, 51, 58, 123
Potato protein and peptides	Antioxidant	Prevent oxidative damage to DNA and other biomolecules.	60, 63, 87
	Antiobesity	Induce CCK release/ enhance response; stimulate CCK1R expression in enteroendocrine cells; inhibit luminal proteases.	65, 71, 92, 96, 98
	Antihyperlipidemic	Increase fecal bile acid and neutral steroid excretion; inhibit cholesterol absorption through suppression of micellar solubility of cholesterol;	46, 122, 128 59, 68, 141

	Antihypertensive	increase cecal short chain fatty acids synthesis. Inhibit ACE	
Potato protease inhibitors	Anticancer Antiobesity	Prevent tumor cell proliferation and H ₂ O ₂ formation; block UV-induced activation of AP-1. Enhance the release of CCK and reduce food intake; enhance response to CCK CCK response primarily by inhibition of trypsin like proteolytic activity	64, 69 65, 70, 71, 92