

¹H NMR (500 MHz, CD₃OD, δ, ppm, J/Hz): 1.00 (3H, s, H-13), 1.04 (3H, s, H-14), 1.82 (3H, s, H-12), 1.90 (3H, s, H-15), 2.17 (1H, d, J = 16.9, H_a-2), 2.48 (1H, d, J = 16.9, H_b-2), 3.17 (1H, dd, J = 8.5, 7.9, H-2'), 3.22–3.35 (3H, m, H-3', 4', 5'), 3.66 (1H, dd, J = 11.9, 5.4, H_a-6'), 3.86 (1H, br.d, J = 11.9, H_b-6'), 4.28 (1H, d, J = 7.9, H-1'), 4.36 (1H, dd, J = 12.8, 7.3, H_a-11), 4.50 (1H, dd, J = 12.8, 6.3, H_b-11), 5.70 (1H, dd, J = 7.3, 6.3, H-10), 5.82 (1H, d, J = 15.9, H-7), 5.89 (1H, s, H-4), 6.36 (1H, d, J = 15.9, H-8). ¹³C NMR (125 MHz, CD₃OD, δ, ppm): 42.7 (C-1), 50.8 (C-2), 201.2 (C-3), 127.2 (C-4), 167.3 (C-5), 80.4 (C-6), 130.0 (C-7), 135.8 (C-8), 137.5 (C-9), 129.4 (C-10), 66.4 (C-11), 13.0 (C-12), 23.5 (C-13), 24.6 (C-14), 19.6 (C-15), 103.3 (C-1'), 75.1 (C-2'), 78.0 (C-3'), 71.7 (C-4'), 78.1 (C-5'), 62.8 (C-6'). FAB-MS *m/z* 435 [M + Na]⁺.

Compound **8** was obtained in trace amounts from a transgenic mutant of *Nicotiana plumbaginifolia* previously, and its structure was assigned tentatively on the basis of MS data without any evidence for the geometry of the double bond [12]. We describe here the NMR data of **8** for the first time, which were assigned by 2D NMR studies including HMBC. The *cis* geometry for the C-9/C-10 double bond was established by NOE experiments. Compound **8**: amorphous solid; [α]_D²⁵ +128° (*c*, 0.17, MeOH). UV (MeOH, λ_{\max} , nm): 234. ¹H NMR (500 MHz, CD₃OD, δ, ppm, J/Hz): 1.00 (3H, s, H-13), 1.05 (3H, s, H-14), 1.88 (3H, s, H-12), 1.91 (3H, s, H-15), 2.20 (1H, d, J = 17.0, H-2a), 2.51 (1H, d, J = 17.0, H_b-2), 3.16 (1H, dd, J = 8.4, 7.9, H-2'), 3.22–3.35 (3H, m, H-3', 4', 5'), 3.68 (1H, dd, J = 11.9, 5.6, H_a-6'), 3.89 (1H, br.d, J = 11.9, H_b-6'), 4.28 (1H, d, J = 7.9, H-1'), 4.36 (1H, dd, J = 12.5, 7.7, H_a-11), 4.49 (1H, dd, J = 12.5, 6.5, H_b-11), 5.61 (1H, dd, J = 7.7, 6.5, H-10), 5.88 (1H, s, H-4), 5.90 (1H, d, J = 15.9, H-7), 6.80 (1H, d, J = 15.9, H-8). ¹³C NMR (125 MHz, CD₃OD, δ, ppm): 42.6 (C-1), 50.7 (C-2), 201.1 (C-3), 127.1 (C-4), 167.1 (C-5), 80.5 (C-6), 132.5 (C-7), 128.2 (C-8), 136.7 (C-9), 127.2 (C-10), 65.4 (C-11), 20.7 (C-12), 23.6 (C-13), 24.8 (C-14), 19.5 (C-15), 103.2 (C-1'), 75.1 (C-2'), 78.0 (C-3'), 71.6 (C-4'), 78.1 (C-5'), 62.8 (C-6'). HR-FAB-MS(+) *m/z* 435.1970 [M + Na]⁺ (C₂₁H₃₂O₈Na requires 435.1995).

Compounds **3–9** were subjected to phytotoxic bioassay against lettuce (*Lactuca sativa*) seed germination using a petri dish method [13]. The compounds were tested at concentrations ranging from 5 to 1000 ppm, and germination was observed daily for five days. (±)-Abscisic acid (purchased from Sigma–Aldrich) was used as a positive control [14, 15].

The phytotoxic activities of **3–9** are expressed as IC₅₀ values: (±)-abscisic acid (5 ppm), **3** (52 ppm), **4** (10 ppm), **5** (5 ppm), **6** (80 ppm), **7** (>500 ppm), **8** (10 ppm), and **9** (>500 ppm). *cis*-Abscisic acid glucoside (**5**) showed the most potent inhibitory activity comparable to (±)-abscisic acid, in accordance with the previous report [8]. *cis*-Abscisic alcohol glucoside (**8**) and vomifoliol (**4**) showed 50% activity against the control. This is the first report to demonstrate the phytotoxic activities of the contents of the fruit of *A. carambola*. None of compounds **3–8** showed antioxidant activity in an assay examining the radical scavenging effect toward DPPH radicals, although the antioxidant activity of (–)-epicatechin (**9**) was confirmed in this study. This study provides evidence that edible fruits are a potential source of environmental friendly bioactive compounds since their safety and toxicological issues related to human beings are remarkably less than those of other natural sources.

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